Characterization of the Western Antarctic Peninsula Ecosystem:
Environmental Controls on the Zooplankton Community

by

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DEDICATION

to my husband, Christian

my daughter, Valentina

and my parents Juan and Marcela
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NOTE TO THE READER

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# TABLE OF CONTENTS

<table>
<thead>
<tr>
<th>Section</th>
<th>Pages</th>
</tr>
</thead>
<tbody>
<tr>
<td>LIST OF TABLES</td>
<td>iv</td>
</tr>
<tr>
<td>LIST OF FIGURES</td>
<td>v</td>
</tr>
<tr>
<td>ABSTRACT</td>
<td>ix</td>
</tr>
<tr>
<td>CHAPTER 1: THE WESTERN ANTARCTIC PENINSULA: PHYSICAL AND BIOLOGICAL SETTING</td>
<td>1</td>
</tr>
<tr>
<td>CHAPTER 2: VALIDATION OF SEAWIFS CHLOROPHYLL A CONCENTRATIONS IN THE SOUTHERN OCEAN: A REVISIT</td>
<td>10-26</td>
</tr>
<tr>
<td>Introduction</td>
<td>10</td>
</tr>
<tr>
<td>Methods</td>
<td>14</td>
</tr>
<tr>
<td>Results</td>
<td>19</td>
</tr>
<tr>
<td>Discussion</td>
<td>26</td>
</tr>
<tr>
<td>CHAPTER 3: SPATIAL AND TEMPORAL VARIABILITY OF SEAWIFS CHLOROPHYLL A DISTRIBUTIONS WEST OF THE ANTARCTIC PENINSULA: IMPLICATIONS FOR KRILL PRODUCTION</td>
<td>30-55</td>
</tr>
<tr>
<td>Introduction</td>
<td>30</td>
</tr>
<tr>
<td>Methods</td>
<td>34</td>
</tr>
<tr>
<td>Results</td>
<td>43</td>
</tr>
<tr>
<td>Discussion</td>
<td>55</td>
</tr>
<tr>
<td>CHAPTER 4: PHYSICAL AND BIOLOGICAL CONTROLS ON INTERANNUAL VARIABILITY OF ZOOPLANKTON IN MARGUERITE BAY, WESTERN ANTARCTIC PENINSULA, AUSTRAL FALL 2001 AND 2002</td>
<td></td>
</tr>
</tbody>
</table>
Introduction ........................................................................................................ 64
Methods ............................................................................................................. 68

Results

Abundance and Percent Contribution ......................................................... 72
Vertical Distribution ...................................................................................... 89
Horizontal Distribution ................................................................................ 96
Fall Environmental Parameters ................................................................. 98
Summer Chlorophyll Concentrations and Krill Recruitment ............... 101

Discussion

Composition and Abundance of Zooplankton in Marguerite Bay .................. 106
Variability in Euphausiid Life History Strategies ...................................... 111
Summer Chlorophyll and Zooplankton Population Response .............. 114
Controls on Zooplankton Spatial Patterns in Marguerite Bay .............. 116
Relationship between Zooplankton and Fall Environmental Parameters ........................................................................................................ 120
Summary ........................................................................................................ 121

CHAPTER 5: SUMMARY AND CONCLUDING REMARKS ......................... 123
REFERENCES CITED .................................................................................. 126

APPENDICES

Appendix 1 ..................................................................................................... 146
Appendix 2 ..................................................................................................... 150
Appendix 3 ..................................................................................................... 154
Appendix 4 ..................................................................................................... 160
### LIST OF TABLES

Table 2.1.  Statistics for the comparisons between \( C_a^{SWF} \) and *in situ* \( C_a \) \((C_a^{FLUOR}, C_a^{HPLC})\) .......................................................... 20

Table 2.2.  Statistics for the comparisons between \( C_a^{FLUOR} \) and \( C_a^{HPLC} \) (mg m\(^{-3}\)) ........ 24

Table 3.1.  Years of elevated krill recruitment between 1975 and 2002 ................. 54

Table 4.1.  Copepod abundance (ind m\(^{-2}\)) in the vicinity of Marguerite Bay during austral fall 2001 and 2002 ................................................................. 74

Table 4.2.  Zooplankton abundance (ind m\(^{-2}\)) in the vicinity of Marguerite Bay during austral fall 2001 and 2002 ................................................................. 75

Table 4.3.  Mean depth of maximum abundance (\( Z, m \)) of copepods in 2001 and 2002 .......................................................................................... 90

Table 4.4.  Mean depth of maximum abundance (m) of euphausiids, amphipods, and mysids during fall 2001 and 2002 ..................................................... 92

Table 4.5.  Spearman rank order correlations between integrated abundance of zooplankton (ind m\(^{-2}\)) and vertically integrated pigment concentrations (chlorophyll + phaeopigment; mg m\(^{-2}\)), salinity at 10 m \((S_{10})\), and bottom depth (bottom \( Z, m \)) ........................................ 100

Table 4.6  Percentage of net hauls in which macrozooplankton were located primarily shallower, deeper, or at the same depth as the thermocline/pycnocline ................................................................. 101
LIST OF FIGURES

Figure 1.1. Map of the western Antarctic Peninsula (WAP) region and geographic references ................................................................. 3

Figure 1.2. General circulation in Marguerite Bay and schematic paths of the Antarctic Circumpolar Current (ACC) (blue) and Antarctic Peninsula Coastal Current (APCC) (red) ................................. 6

Figure 1.3. Bathymetric map of Marguerite Bay and adjacent waters of the western Antarctic Peninsula ......................................................... 7

Figure 2.1. Sampling stations overlaid on SeaWiFS images of mean $C_a$ for January (a) 1998, (b) 1999, (c) 2000, (d) 2001 and (e) 2002 ...................... 18

Figure 2.2. Distribution of $in situ$ depth-weighted (a) $C_a^{Fluor}$ and (b) $C_a^{HPLC}$ during January-February 1999 ..................................................... 19

Figure 2.3. Comparison between $C_a^{SWF}$ (mg m$^{-3}$, SeaDAS4.8, OC4v4 algorithm) and $in situ$ $C_a$ (mg m$^{-3}$) .......................................................... 22

Figure 2.4. Comparison between $C_a$ predicted by the OC4v4 algorithm (using SeaWiFS-derived $R_{rs}$ as input) and measured $in situ$ $C_a$ (mg m$^{-3}$) .......... 23

Figure 2.5. Comparison between $C_a^{HPLC}$ and $C_a^{Fluor}$ (mg m$^{-3}$) between January and February 1998 – 2001 ...................................................... 25

Figure 2.6. Normalized histogram of $C_a^{SWF}$ distributions (mg m$^{-3}$) in the Southern Ocean during austral summer ........................................ 26

Figure 2.7. Relationship between HPLC $C_b/C_a$ and $C_a^{Fluor}/C_a^{HPLC}$ and between HPLC $C_c/C_a$ and $C_a^{Fluor}/C_a^{HPLC}$ ........................................ 29
Figure 3.1. Map of the Antarctic continent showing the location of the study area and other geographic references ............................................. 37

Figure 3.2. Mean abundance (ind m$^{-3}$) of development stages of *Euphausia superba* in offshelf waters (Sta. 1) west of the Antarctic Peninsula and coastal waters (Sta. 5) of Marguerite Bay during fall 2001 (light grey) and 2002 (dark grey) ......................................................... 41

Figure 3.3. Biweekly climatology (1997 - 2004) of SeaWiFS chlorophyll concentrations (mg m$^{-3}$) between October and March ...................... 44

Figure 3.4a. Location of the 14 subregions along the western Antarctic Peninsula superimposed over the climatology (1998 - 2004) of SeaWiFS chlorophyll concentrations (mg m$^{-3}$) for January 1 - 14 ......................... 47

Figure 3.4b. Time series of geometric mean chlorophyll concentrations in each subregion for each biweekly period during 1997 - 2004 ..................... 48

Figure 3.5a. Biweekly SeaWiFS chlorophyll concentrations (Chl, mg m$^{-3}$) in October (Oct), November (Nov) and December (Dec) of 2000 ........... 49

Figure 3.5b. Biweekly SeaWiFS chlorophyll concentrations (Chl, mg m$^{-3}$) in October (Oct), November (Nov) and December (Dec) of 2001 .......... 50

Figure 3.6. Biweekly time series of chlorophyll accumulation after the ice had receded at the September, October, and November 2000 and 2001 locations of the ice edge shown in figure 3.5 .................................................. 52

Figure 3.7. Daily ice-free area (km$^{2}$) in (a) northern and (b) southern Marguerite Bay during 1997 - 2004 ......................................................... 53

Figure 3.8. Monthly mean SeaWiFS chlorophyll concentrations in the Bellingshausen Sea during February (a) 2001 and (b) 2002 ............... 63
Figure 4.1.  (a) Location of the study area (red rectangle) and MOCNESS net hauls (circles) during fall of (b) 2001 and (c) 2002. 70

Figure 4.2a.  Water column integrated abundance of copepods (ind m$^{-2}$) from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 76

Figure 4.2b.  Percent composition of copepods at coastal stations in Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 77

Figure 4.3a.  Water column integrated abundance of euphausiids (ind m$^{-2}$) from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 79

Figure 4.3b.  Percent composition of euphausiids from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 80

Figure 4.4.  Length frequency of *E. superba* juveniles and adults during fall 2001 (top) and 2002 (bottom) 84

Figure 4.5 Length frequency of *E. superba* juveniles and adults during fall 2001 (top) and 2002 (bottom) 85

Figure 4.6 Length frequency of *E. crystallorophias* juveniles and adults during fall 2001 (top) and 2002 (bottom) 86

Figure 4.7a.  Water column integrated abundances of zooplankton other than copepods and euphausiids (ind m$^{-2}$) from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 87

Figure 4.7b.  Percent composition of zooplankton from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom) 88
Figure 4.8. Mean depth of maximum abundance (m) of copepods during 2001 (black) and 2002 (grey) ................................................................. 91

Figure 4.9a. Vertical distribution of euphausiids, amphipods and mysids (ind m$^{-3}$) in Crystal Sound during austral fall 2001 (top) and 2002 (bottom) ................................................................. 93

Figure 4.9b. Vertical distribution of euphausiids, amphipods and mysids (ind m$^{-3}$) in Laubeuf Fjord during austral fall 2001 (top) and 2002 (bottom) ................................................................. 94

Figure 4.9c. Vertical distribution of euphausiids, amphipods and mysids (ind m$^{-3}$) in the vicinity of Alexander Island during austral fall 2001 (top) and south of Adelaide Island in fall 2002 (bottom) ................. 95

Figure 4.10a. Linear correlation between integrated abundances (ind m$^{-2}$) of total macrozooplankton (euphausiids, amphipods, and mysids) and copepods at different stations during fall 2001 and 2002 (n = 12; r = -0.39; p = 0.208) ................................................................. 96

Figure 4.10b. Mean water-column integrated abundance (ind m$^{-2}$) of macrozooplankton (euphausiids, amphipods and mysids) (ind m$^{-2}$) (grey bars, left axis) and copepods (black circles, right axis) at different stations within Marguerite Bay during fall 2001 (top) and 2002 (bottom) ................................................................. 97

Figure 4.11. (a) Location of the subregions analyzed for median SeaWiFS chlorophyll concentrations (mg m$^{-3}$) in (b, c) oceanic and (d) coastal waters of the Bellingshausen Sea, (e) northern, and (f) southern Marguerite Bay in spring/summer 2000/2001 (grey) and 2001/2002 (black) ................................................................. 103

Figure 4.12. (b) Spearman R correlation between geometric mean chlorophyll concentrations in oceanic waters of the Bellingshausen Sea during November 1997 - 2004 and recruitment of $E. superba$ ($R_1$) in waters west of the Antarctic Peninsula; n = 6; R = 0.81; p < 0.05) ................................................................. 105
Characterization of the Western Antarctic Peninsula Ecosystem: Environmental Controls on the Zooplankton Community

Marina Marrari

ABSTRACT

The zooplankton community of Marguerite Bay, western Antarctic Peninsula, was investigated in relation to variability in chlorophyll concentrations and sea ice dynamics, using a combination of satellite remote sensing techniques and plankton net data. SeaWiFS chlorophyll data were validated with concurrent in situ data measured by HPLC and fluorometric methods, and results indicate that SeaWiFS chlorophyll is an accurate measure of in situ values when HPLC data are used as ground truth.

Climatology data of SeaWiFS chlorophyll west of the Antarctic Peninsula showed that the Bellingshausen Sea and Marguerite Bay usually had higher and more persistent chlorophyll concentrations compared with northern regions. These predictable phytoplankton blooms could provide the Antarctic krill, Euphausia superba, with the food required for successful reproduction and larval survival. Unusually high krill reproduction in 2000/2001 was coincident with above-average chlorophyll concentrations throughout the study area and was followed by the largest juvenile recruitment since 1981. High larval densities at the shelf break along the Antarctic Peninsula may have
resulted, in part, from krill spawning in the Bellingshausen Sea. Interannual differences in sea ice also probably contributed to the variability in larval krill abundances.

Interannual differences were observed in the species composition of the zooplankton of Marguerite Bay during fall, and these were linked to variability in the environmental conditions. *Thysanoessa macrura* was the most abundant euphausiid in 2001, while *Euphausia crystallorophias* dominated in 2002, and *E. superba* had intermediate densities during both years. Copepods were more abundant in 2001 by a factor of 2.6. Copepods and *T. macrura* showed a rapid population response to unusually high chlorophyll concentrations in the Bellingshausen Sea and Marguerite Bay during spring-summer 2000/2001, whereas *E. superba* and *E. crystallorophias* had a longer term response and showed increased recruitment in fall 2002. There were no clear associations between the distribution of zooplankton and environmental conditions in fall; however there was a significant relationship between chlorophyll concentrations in the Bellingshausen Sea during the preceding spring and zooplankton patterns during fall.
CHAPTER ONE

THE WESTERN ANTARCTIC PENINSULA: PHYSICAL AND BIOLOGICAL SETTING

The Southern Ocean covers approximately 10% of the world’s ocean, supporting large concentrations of zooplankton and higher trophic level organisms. The region plays an important role in the world’s ocean biogeochemical cycle (Sarmiento et al., 1998) and has a profound influence on global circulation by connecting the Atlantic, Pacific, and Indian Ocean basins, and through deep water formation. Based on biogeography, three main zones have been described for the Southern Ocean: (1) an ice-free zone rich in nutrients, but relatively poor in primary production, with a zooplankton community dominated by copepods, salps and small euphausiids, (2) a productive seasonal pack-ice zone that includes the area covered by sea ice during winter, but mostly ice-free during summer and fall, where zooplankton may be dominated by the Antarctic krill, *Euphausia superba*, and (3) a permanent pack ice zone, with generally low zooplankton biomass, where *E. superba* is often replaced by the neritic euphausiid, *Euphausia crystallorophias* (Hempel, 1985).

Within the Atlantic sector of the Southern Ocean, the continental shelf along the western Antarctic Peninsula (WAP) includes waters of the seasonal and permanent pack ice zones (Fig. 1.1). This region supports high concentrations of zooplankton and
predators, and is considered one of the most productive areas in Antarctic waters (Deibel and Daly, 2007). The Antarctic Peninsula region also is of interest as it is warming more rapidly than almost any other place on the planet. A considerable increase in atmospheric and upper-ocean temperatures has occurred in the last 50 years, while ice shelves have retreated (King, 1994; Smith et al., 1996b; Vaughan and Doake, 1996; Vaughan et al., 2003). In addition, there have been marked changes in winter sea ice extent and duration (Parkinson 2002). These changes have important implications for Antarctic organisms that rely directly on sea ice for reproduction, such as Adèlie penguins (Trivelpiece and Fraser, 1996), as well as for others that will suffer indirect effects through changes in prey populations (Costa and Crocker, 1996).

The Antarctic krill, *E. superba*, is a relatively large pelagic crustacean (up to 65 mm in length) and a keystone species in the Antarctic ecosystem, acting as a primary grazer on phytoplankton and prey for a variety of higher trophic level predators. Most Antarctic predators, including species of fish, seals, whales, penguins, and many seabirds, rely on zooplankton for survival, and in particular Antarctic krill (Lowry et al., 1988; Ainley and DeMaster, 1990; Costa and Crocker, 1996; Murase et al., 2002). *E. superba* has a circumpolar distribution, with highest concentrations in the Atlantic sector of the Southern Ocean (Marr, 1962). It is believed that krill originated in the WAP region are the source of large populations observed downstream in the Scotia Sea and at South Georgia Island (Fach and Klinck, 2006; Thorpe et al., 2007).

Since the *Discovery* expeditions in the 1920-30s, which surveyed almost the entire Southern Ocean and resulted in a large amount of information regarding the distribution, biology and ecology of Antarctic krill (Marr, 1962), research in the vicinity
of the WAP has been mostly restricted to northern waters along the continental shelf and downstream in the Scotia Sea.

Figure 1.1. Map of the western Antarctic Peninsula (WAP) region and geographic references. The dotted line indicates the 1000 m isobath.

Some interdisciplinary programs that investigated the structure and dynamics of the marine ecosystem from the mid- to northern sectors of the WAP include BIOMASS Program (Biological Investigations of Marine Antarctic Systems and Stocks) in 1985/86
(El-Sayed, 1994), RACER (Research on Antarctic Coastal Ecosystem Rates) (Huntley et al., 1991), Palmer LTER (Long Term Ecological Research), which since the early 1990s has been dedicated to studying how physical forcing affects the structure and function of the ecosystem (Ross et al., 1996), and AMLR (Antarctic Marine Living Resources Program), established in 1990 to assess resources in the area around Elephant Island (e.g., Loeb et al., 1997; Siegel et al., 2002; Watkins et al., 2004). The information resulting from these efforts, as well as from other field expeditions from the United Kingdom, Germany, Argentina, Poland, Russia, and Chile, among others, has greatly improved our understanding of the processes controlling populations in the area, and has provided insight into the dominant physical components, such as circulation, hydrography, nutrient distributions, and sea ice dynamics. However, in spite of the significant amount of information available for the northern sectors, there have been few studies in southern regions of the WAP (Marguerite Bay and the eastern Bellingshausen Sea) (Atkinson, 1995; Siegel and Harm, 1996; Lascara et al., 1999; Meyer et al., 2003).

The Southern Ocean Global Ocean Ecosystem Dynamics Program (SOGLOBEC) field efforts were focused in the vicinity of Marguerite Bay, as limited information suggested that it was an important overwintering site for upper trophic level predators and, therefore, probably of their food source, Antarctic krill (Hoffman et al., 2004). The primary goal of the U.S. SOGLOBEC Program was to investigate the physical and biological factors that influence the growth, recruitment, and overwintering survival of *E. superba* in the vicinity of Marguerite Bay as well as the associated predators and prey of krill (Hoffman et al., 2004).

Initial results of the US SOGLOBEC Program have described the general
circulation in Marguerite Bay. There are two major currents: the wind-driven westerly
Antarctic Circumpolar Current (ACC) that flows northward along the continental shelf
break, and the buoyancy-driven Antarctic Peninsula Coastal Current (APCC), which
flows southward along the coast (Moffat et al., 2008) (Fig. 1.2). The APCC enters
Marguerite Bay at the south end of Adelaide Island, flows clockwise along the coast and
exits around the north end of Alexander Island. Mesoscale circulation features, such as
gyres on the outer shelf and eddies within inner Marguerite Bay, contribute to making
Marguerite Bay a favorable retention area for phyto- and zooplankton (Beardsley et al.,
2004; Klinck et al., 2004; Dorland and Zhou, 2008)

Marguerite Bay encompasses a relatively wide and deep continental shelf, with
mean depths of ~ 400 m (Bolmer et al., 2004). The bathymetry is influenced by
Marguerite Trough, a deep canyon up to 1600 m deep, which intersects the continental
shelf off Marguerite Bay, crosses the shelf, and extends into George VI Sound (Fig. 1.3).
In addition, the area is characterized by numerous seamounts and depressions that result
in a variable and complicated topography, particularly in the northern sectors, such as
Laubeuf Fjord and Crystal Sound north of Marguerite Bay. Intrusions of Upper
Circumpolar Deep Water (UCDW), found at depths greater than 500 m in oceanic areas,
have been observed on the continental shelf, supplying Marguerite Bay with warm and
nutrient rich waters at depth. These intrusions occur through Marguerite Trough, as well
as in other shelf break areas such as those off Crystal Sound and west of Alexander Island
(Fig. 1.2) (Dinniman and Klinck, 2004).

The hydrographic structure of continental shelf waters along the Antarctic Peninsula has
been previously described (Smith et al., 1999; Klinck et al., 2004). The primary water
masses include Antarctic Surface Water (ASSW), characterized by temperatures and salinity ranges between -1.8 and 1 °C and 33.0 and 33.7, respectively. Beneath a pycnocline, generally established at ~ 100 - 120 m, is warm (1.5 °C) and salty (34.6 – 34.73) water derived from Circumpolar Deep Water (CDW), which is typically present in waters over the outer shelf of the WAP at depths of 200 - 600 m. During late fall, surface
heat loss and wind forcing result in a deep winter mixed layer occupied by near-freezing (< -1 °C) Winter Water (WW). This surface layer is partially restratified in spring and a remnant layer of cold WW is normally observed at ~ 80 - 100 m between the AASW and modified CDW layers.

Figure 1.3. Bathymetric map of Marguerite Bay and adjacent waters of the western Antarctic Peninsula. Bathymetry in meters (m). Reproduced from Bolmer et al. (2004).

A major physical characteristic of the Southern Ocean is the annual advance and retreat of sea ice. During summer most of the WAP continental shelf is ice-free, allowing dense phytoplankton blooms to develop. During winter, sea ice cover can
completely cover the continental shelf. In recent years, however, a dramatic decline in sea
ice extent has been observed in the WAP, resulting in areas of the northern WAP which
may remain ice-free throughout the year. However, in southern sectors, such as
Marguerite Bay and the eastern Bellingshausen Sea, winter sea ice generally extends over
most of the continental shelf and, thus, can be considered an important factor influencing
marine organisms. Extensive sea ice reduces the penetration of radiant heat into the
water column during spring, summer, and fall, limiting phytoplankton growth and, thus,
food available to zooplankton.

To summarize, the Marguerite Bay region is characterized by a complex
seafloor topography, extreme variability in seasonal sea ice conditions, and circulation
features, which include opposite flowing currents, intrusions of oceanic waters onto the
shelf, and mesoscale gyres and eddies. Given the rapidly changing conditions of this
region and the potential effects on Antarctic organisms, understanding zooplankton
population dynamics and their response to environmental variability is critical to
predicting the effects of climate change on the Antarctic ecosystem as a whole.

This dissertation aims to investigate the effects of environmental variability on
the population dynamics of phytoplankton and zooplankton in the vicinity of Marguerite
Bay, with special emphasis on *E. superba*. For this purpose, I use a combination of ocean
color remote sensing techniques, zooplankton net data, and environmental information.
The main objective of Chapter 2 is to evaluate the performance of the SeaWiFS satellite
sensor (Sea-Viewing Wide Field-of-View Sensor) in estimating chlorophyll
concentrations in the Southern Ocean. Chapter 3 focuses on using SeaWiFS data to
describe the temporal and spatial dynamics of chlorophyll distributions along the WAP
during spring and summer, and relate the interannual variability observed in chlorophyll
dynamics to changes in the reproduction and recruitment success of *E. superba*. Chapter
4 investigates the patterns of abundance and distribution of the dominant zooplankton
groups in Marguerite Bay, in relation to variability in environmental conditions, and a
summary of the major findings and future direction is presented in Chapter 5.
CHAPTER TWO

VALIDATION OF SEAWIFS CHLOROPHYLL A CONCENTRATIONS IN THE SOUTHERN OCEAN: A REVISIT

INTRODUCTION

Since the launch of the Sea-viewing Wide Field-of-view Sensor (SeaWiFS, McClain et al., 1998) onboard the Orbview-II satellite in August 1997, ocean color data products, in particular concentrations of chlorophyll \( a (C_a, \text{mg m}^{-3}) \) in the surface ocean, have been used to investigate a wide variety of fundamental topics including ocean primary productivity, biogeochemistry, coastal upwelling, eutrophication, and harmful algal blooms (e.g., Hu et al., 2005; Muller-Karger et al., 2004). Other ocean color missions, such as the ongoing MODerate-resolution Imaging Spectroradiometer (MODIS, Esaias et al., 1998; Terra satellite for morning pass since 1999 and Aqua satellite for afternoon pass since 2002) or the future National Polar-Orbiting Operational Environmental Satellite System (NPOESS), assure the continuity of remotely sensed ocean color in assessing the long-term global change in several key environmental parameters, including \( C_a \). Quantitative use of ocean color data products requires a high level of accuracy. During algorithm development, the errors in the \( C_a \) data products after logarithmic transformation were about 0.2 or less (O’Reilly et al., 2000), which corresponds to roughly 50% root mean square (RMS) relative error. Global validation efforts show that in most ocean basins \( C_a \) errors are about 0.3 (Gregg and Casey, 2004)
although in regions such as the Southern Ocean, reported errors are significantly larger.

The Southern Ocean was defined by the International Hydrographic Organization in 2000 to encompass waters between the northern coast of Antarctica and 60° S. Oceanographers, however, traditionally have defined the northern limit of the SO as the Subtropical Front (at approximately 40° S) (Orsi et al., 1995). Typical chlorophyll concentrations in the Southern Ocean range between 0.05 and 1.5 mg m⁻³ (Arrigo et al., 1998; El-Sayed, 2005). It is believed that the interaction of light and deep mixing, iron, and grazing limit phytoplankton growth throughout the Southern Ocean, in addition to low silicate concentrations which can limit diatom production north of the Polar Front (Moline and Prézelin, 1996; Daly et al., 2001; Boyd, 2002). However, elevated chlorophyll concentrations (1 to > 30 mg m⁻³) are characteristic of many regions, including continental shelf and ice edge areas (Holm-Hansen et al., 1989; Moore and Abbott, 2000; El-Sayed, 2005), and even values of up to 190 mg m⁻³ have been reported (El-Sayed, 1971). The Antarctic Peninsula region, in particular, supports large concentrations of phytoplankton, zooplankton, seabirds, seals and whales, and is considered one of the most productive areas of the Southern Ocean, for reasons that are not fully understood (Deibel and Daly, 2007).

Several studies have relied on ocean color data to investigate phytoplankton spatial patterns (Moore and Abbott, 2000; Holm-Hansen et al., 2004a), interannual variability during summer (Smith et al., 1998a; Korb et al., 2004) and primary productivity (Dierssen et al., 2000; Smith et al., 2001) west of the Antarctic Peninsula and in the adjoining Scotia Sea. These studies used $in situ$ $C_a$ determined from water samples using fluorometric methods ($C_a^{\text{Fluor}}$) to validate monthly/weekly averages of
SeaWiFS \( C_a (C_a^{SWF}) \) data product at \( \sim 9 \times 9 \text{ km}^2 \) or \( \sim 4 \times 4 \text{ km}^2 \) resolution and concluded that in the Southern Ocean, \( C_a^{SWF} \) values are significantly lower than those estimated from \textit{in situ} water samples. For example, Dierssen and Smith (2000) applied \textit{in situ} bio-optical data measured between 1991 and 1998 to the OC2v2 algorithm to test its applicability west of the Antarctic Peninsula in the Southern Ocean. They concluded that \( C_a \) derived from the OC2v2 algorithm using \textit{in situ} reflectance was 60% lower than \textit{in situ} \( C_a \) (\( C_a \) between 0.7 and 43 mg m\(^{-3}\), median \( \sim 1 \text{ mg m}^{-3} \)). Korb et al. (2004) reported that \( C_a^{SWF} \) values were only 87% of \( C_a^{Fluor} \) for concentrations lower than 1 mg m\(^{-3}\) and only 30% for concentrations above 5 mg m\(^{-3}\) in the South Georgia area (54.5º S, 37º W).

In addition, Moore et al. (1999) found a strong linear relationship between \( C_a^{SWF} \) and \( C_a^{Fluor} \) (\( R^2 = 0.72, n = 84 \)) in the Ross Sea, although they noted that SeaWiFS tended to underestimate \( C_a \) values between 0.1 and 1.5 mg m\(^{-3}\).

The previous validation methods may present several limitations. First, \textit{in situ} samples are point measurements while satellite pixels cover a larger area (up to \( 9 \times 9 \text{ km}^2 \)). Patchiness within a pixel will affect the comparison of results between areas and over time (e.g., Hu et al., 2004). Second, the \textit{in situ} and satellite measurements are not strictly concurrent and the time differences can be large (up to a month). Finally, and most importantly, previous validation studies used \textit{in situ} \( C_a \) from fluorometric measurements, while it is now widely recognized that High Performance Liquid Chromatography (HPLC) may yield more accurate results in determining \( C_a \) from water samples. Fluorometric methods may result in biased results, particularly in the presence of certain accessory pigments (Lorenzen, 1981; Welschmeyer, 1994).

In a study that included three different areas of the world’s oceans, Trees et al.
(1985) reported that errors in the $C_a^{\text{Fluor}}$ ranged between -68 and 53% with a mean of 39%. In addition, Bianchi et al. (1995) found that $C_a^{\text{Fluor}}$ in the northern Gulf of Mexico was approximately 30% lower than $C_a^{\text{HPLC}}$, except in near coastal areas. It is believed that the presence of significant amounts of chlorophyll b ($C_b$), characteristic of chlorophytes, prochlorophytes and prasinophytes, causes fluorometric techniques to underestimate $C_a$. On the other hand, high concentrations of chlorophyll c ($C_c$), typically found in diatoms, dinoflagellates, cryptophytes, and haptophytes, lead to an overestimation of $C_a$ with respect to fluorometric measurements. The fluorescence emission spectra of degradation products (phaeopigments) of $C_a$ and $C_b$ overlap considerably, causing an overestimation of $C_a$ phaeopigments and, thus, an underestimation of $C_a$. On the other hand, $C_a$ and $C_c$ have partially overlapping fluorescence spectra, causing an overestimation of $C_a$ and subsequent underestimation of phaeopigments $a$ (Gibbs, 1979; Jeffrey et al., 1997). The filters used in the standard fluorometric method (Lorenzen, 1981) cannot effectively discriminate between $C_a$, $C_b$, $C_c$ and their degradation products; thus, depending on the type of phytoplankton present and their associated pigments, $C_a$ may be overestimated or underestimated by fluorometric methods.

Herein, I use concurrent HPLC and fluorometric data collected between 1998 and 2002 in waters west of the Antarctic Peninsula, as well as high-resolution SeaWiFS data, to re-examine whether SeaWiFS $C_a$ is underestimated in the Southern Ocean as reported in previous studies. I also discuss possible explanations for the observed results and investigate the effects of different accessory pigments on $C_a$ estimations.
METHODS

SeaWiFS daily Level 2 data between December 1997 and December 2004 were obtained from NASA Goddard Space Flight Center. These data were derived from the high-resolution (~ 1 km/pixel near nadir) Level 1 data collected by ground stations, as well as occasional satellite onboard recording over the area using the most current algorithms and software package (SeaDAS4.8). A total of 6606 data files were obtained and mapped to a rectangular projection with approximately 1 km²/pixel for the area between 45° - 75° S and 50° - 80° W west of the Antarctic Peninsula (Fig. 1.1). The data product used in this study is the surface $C_a$ estimated with the OC4v4 empirical algorithm (O’Reilly, 2000):

$$C_a = 10^{0.366 - 3.067R + 1.93R^2 + 0.649R^3 - 1.532R^4}$$

where $R = \log_{10}[(\max(R_{rs443}, R_{rs490}, R_{rs510}))/R_{rs555})]$ and $R_{rs}$ is the remote sensing reflectance, a data product after atmospheric correction.

Chlorophyll fluorescence and HPLC pigment data were collected and analyzed by Drs Raymond Smith (University of California Santa Barbara) and Maria Vernet (University of California San Diego) as part of the Palmer Long Term Ecological Research (LTER) program during cruises west of the Antarctic Peninsula (see http://pal.lternet.edu/data/ for detailed methods). The location of the LTER chlorophyll sampling stations between 1998 and 2002 are shown in figure 2.1. Most of the samples were collected within the 2000 m isobath, although two transects were conducted across
Drake Passage in January-February 1999 and 2000 to measure \( C_a^{\text{Fluor}} \). At each station, water column samples were collected at discrete depths for both fluorometric and HPLC measurements. \( C_a, C_b \) and \( C_c \) were obtained by HPLC from samples collected at fixed stations during January-February 1998 and 1999 following the methods of Wright et al. (1991), and during January-February 2000 and 2001 following the methods of Zapata et al. (2000).

\( C_a \) and phaeopigment concentrations also were obtained by fluorometric methods by measuring total fluorescence and subtracting phaeopigments after acidification from samples collected during January - February 1998, 1999, 2000, 2001 and 2002 following Smith et al. (1981, 1996a, 1998). Welschmeyer’s (1994) method, which effectively measures fluorescence from \( C_a \) only and reduces interference from \( C_b \) or its phaeo-derivatives, was not applied (M. Vernet, pers. comm.).

Because the signal detected by the satellite sensor is an optically-weighted function of signals at all depths (up to 50 - 60 m for clear waters), the method of Gordon (1992) was used to calculate a depth-weighted chlorophyll concentration, \( <C> \), to compare with satellite estimates:

\[
< C > = \frac{\int_0^z g(z') C(z') dz'}{\int_0^z g(z') dz'}
\]  

(2)

where \( g(z) = \exp[-2\int_0^z K(z') dz'] \) and \( z \) is the depth. \( K \) is the diffuse attenuation coefficient that is approximated by \( K (z) \approx 0.121 C(z)^{0.428} \) (Morel, 1988). The integration was from 0 to 50 m and included 5 or 6 vertical samples at most stations, although in some cases only 3 - 4 samples were available for the calculations. A total of 189 HPLC
and 775 fluorometric $C_a$ values were used in these analyses. Because the weighting function, $g(z)$, decreases exponentially with increasing depth, $<C>$ is not very different from the surface value, at least for fluorometric $C_a$ (ratio = 1.02 ± 0.15, p = 0.841). For the HPLC samples, the differences between $<C>$ and surface $C_a$ are significant (ratio = 1.05 ± 0.99, p = 0.022). The daily, high-resolution SeaWiFS $C_a$ data were queried to compare with the in situ data in the following manner. To reduce errors caused by digitization and random noise, for each in situ data point, all valid satellite data from a 5 x 5 pixel box covering the in situ location (except those cloud and land adjacent pixels) were used to compute the median value (Hu et al., 2001). A rigorous comparison between satellite and in situ data should limit the time difference between the two measurements to within ±2 - 3 hours. Due to extended cloud coverage and the occasional presence of sea ice, however, only a small number of HPLC data points were obtained under such rigorous criteria, leading to statistically meaningless results. Therefore, the time difference between satellite and in situ measurements was relaxed to ±3 days.

Estimating uncertainty in a satellite-derived parameter with log-normal distribution is not trivial, as discussed in Campbell (submitted). Here, two estimates were used to assess the differences between the in situ and satellite-derived data. First, the root mean square (RMS) and the mean difference (bias) in percentage were defined as:
\[ RMS = \sqrt{\frac{1}{n} \sum_{i=1}^{n} (x_i)^2} \times 100 \]

\[ bias = \bar{x} = \left( \frac{1}{n} \sum_{i=1}^{n} x_i \right) \times 100 \]  

\[ x = \frac{S - I}{I} \]  

where \( S \) is satellite data, \( I \) is in situ data, and \( n \) is the number of data pairs. For a normally distributed \( x \), RMS should equal the standard deviation. Further, because the natural distribution of \( C_a \) is lognormal (Campbell, 1995), error estimates were also made on the logarithmically transformed (base 10) data:

\[ \text{log}_{\text{RMS}} = \sqrt{\frac{\sum [(\log(S) - \log(I))^2]}{n}} \]

\[ \text{log}_{\text{bias}} = \frac{\sum [\log(S) - \log(I)]}{n} \]  

These error estimates have been used in recent publications to describe the performance of the ocean color algorithms (O’Reilly et al., 2000) and to validate SeaWiFS global and regional estimates of \( C_a \) (Darecki and Stramski, 2004; Gregg and Casey, 2004; Zhang et al., 2006). Note that these latter error estimates cannot be expressed as percentages because they are logarithmically transformed (Campbell, submitted).
Figure 2.1. Sampling stations overlaid on SeaWiFS images of mean $C_d$ for January (a) 1998, (b) 1999, (c) 2000, (d) 2001 and (e) 2002. White circles: fluorometric samples, pink triangles: HPLC samples, white line: 2000 m isobath.
RESULTS

Typical $C_a^{\text{Fluor}}$ and $C_a^{\text{HPLC}}$ distributions during austral summer are presented for January-February 1999 (Fig. 2.2). In all years, $C_a^{\text{Fluor}}$ ranged from 0.052 to 27.6 mg m$^{-3}$, with a median of 0.86 mg m$^{-3}$. $C_a^{\text{HPLC}}$ was typically lower and ranged from 0.017 to 14.6 mg m$^{-3}$ with a median of 1.04 mg m$^{-3}$. In general, the lowest $C_a$ values (<0.1 mg m$^{-3}$) were consistently found offshelf in Drake Passage. Elevated $C_a$ values (>1 mg m$^{-3}$) were detected throughout the continental shelf, with the highest values (>10 mg m$^{-3}$) always observed in Marguerite Bay.

![Figure 2.2](image)

Figure 2.2. Distribution of in situ depth-weighted (a) $C_a^{\text{Fluor}}$ and (b) $C_a^{\text{HPLC}}$ during January-February 1999. White line: 2000 m isobath.

A total of 96 $C_a^{\text{SWF}}$-$C_a^{\text{HPLC}}$ matching pairs and 307 $C_a^{\text{SWF}}$-$C_a^{\text{Fluor}}$ matching pairs were obtained using the method described above. Table 2.1 lists the statistics of these comparisons. In general, $C_a^{\text{SWF}}$ is significantly lower than $C_a^{\text{Fluor}}$ (Fig. 2.3), with a ratio of 0.55 ± 0.63 between the two (Table 2.1). The inverse ratio, i.e., the ratio of $C_a^{\text{Fluor}}/C_a^{\text{SWF}}$, is 2.73 ± 2.19, consistent with previous observations in the Southern Ocean.
where $C_{a}^{\text{Fluor}}$ was used to validate $C_{a}^{\text{SWF}}$ and the same pattern of underestimation was observed (Moore et al., 1999; Dierssen and Smith, 2000; Korb et al., 2004). In contrast, $C_{a}^{\text{HPLC}}$ showed a more satisfactory agreement with $C_{a}^{\text{SWF}}$ over a wide dynamic range (0.1 – 4 mg m$^{-3}$) (Fig. 2.3). The mean ratio of $C_{a}^{\text{SWF}} / C_{a}^{\text{HPLC}}$ is close to 1 (i.e. 1.12), in contrast to the lower ratio of 0.55 for $C_{a}^{\text{SWF}} / C_{a}^{\text{Fluor}}$.

Table 2.1. Statistics for the comparisons between $C_{a}^{\text{SWF}}$ and in situ $C_{a}$ ($C_{a}^{\text{Fluor}}$, $C_{a}^{\text{HPLC}}$). $n$ is the number of matching pairs, RMS is root mean square error, SD is standard deviation.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>$C_{a}^{\text{SWF}}$ vs. $C_{a}^{\text{Fluor}}$</th>
<th>$C_{a}^{\text{SWF}}$ vs. $C_{a}^{\text{HPLC}}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>$n$</td>
<td>307</td>
<td>96</td>
</tr>
<tr>
<td>Ratio ± SD</td>
<td>0.55 ± 0.63</td>
<td>1.12 ± 0.91</td>
</tr>
<tr>
<td>RMS</td>
<td>77.2%</td>
<td>91.4%</td>
</tr>
<tr>
<td>Bias</td>
<td>-45.2%</td>
<td>12%</td>
</tr>
<tr>
<td>log RMS</td>
<td>0.44</td>
<td>0.34</td>
</tr>
<tr>
<td>log bias</td>
<td>-0.36</td>
<td>-0.07</td>
</tr>
</tbody>
</table>

Although the RMS errors for the two comparisons are comparable (Table 2.1), $C_{a}^{\text{HPLC}}$ is nearly equally scattered around the 1:1 line (Fig. 2.3), suggesting that the bias errors in $C_{a}^{\text{SWF}} / C_{a}^{\text{HPLC}}$ are significantly smaller than those in $C_{a}^{\text{SWF}} / C_{a}^{\text{Fluor}}$. Clearly, the agreement between $C_{a}^{\text{SWF}}$ and $C_{a}^{\text{HPLC}}$ is much improved over that between $C_{a}^{\text{SWF}}$ and $C_{a}^{\text{Fluor}}$.

Similar results were also obtained from the algorithm perspective. By using the spectral remote sensing reflectance data ($R_{n}$) derived from satellite measurements (Fig. 2.4), the OC4v4 algorithm yielded comparable results to those obtained from HPLC measurements. In contrast, $C_{a}^{\text{Fluor}}$ values are significantly higher than those predicted by the OC4v4 algorithm for the entire range considered.
Are these results representative of the entire Southern Ocean? Due to cloud cover, satellite data were not available for all pixels every day. This reduced the number of $C_a^{SWF}$ data points, which resulted in a limited number of matching pairs for comparing satellite and in situ data (307 for fluorometric and 96 for HPLC). However, the in situ data itself comprised a much larger dataset that included 832 concurrent fluorometric and HPLC measurements. When this in situ dataset was used to compare $C_a^{Fluor}$ and $C_a^{HPLC}$, similar results were obtained, i.e., the mean ratio of $C_a^{Fluor}/C_a^{HPLC}$ is 2.43 ± 3.37 (Fig. 2.5). The ratio of $C_a^{Fluor}/C_a^{HPLC}$ appears to decrease with increasing concentrations (Table 2.2), although for $C_a^{HPLC} <0.05 \text{ mg m}^{-3}$ and $C_a^{HPLC} >3.0 \text{ mg m}^{-3}$ the statistical results may not be reliable because of the few matching pairs available and the scatter of the data (Fig. 2.5). For $C_a^{HPLC}$ between 1.5 and 3.0 mg m$^{-3}$, the bias is small (15%) and the mean ratio of $C_a^{Fluor}/C_a^{HPLC}$ is close to unity (1.15 ± 0.73). Between 0.05 and 1.5 mg m$^{-3}$, however, $C_a^{Fluor}$ is much higher than $C_a^{HPLC}$ (mean $C_a^{Fluor}/C_a^{HPLC} = 2.48 \pm 2.23$, n = 647). This difference is believed to be due to errors in the $C_a^{Fluor}$ measurements as described above. Because most (> 90%) of the waters in the Southern Ocean have surface $C_a^{SWF}$ values between 0.05 and 1.5 mg m$^{-3}$ (Fig. 2.6), this assessment can be generalized and applied to most regions.
Figure 2.3. Comparison between $C_{a}^{SWF}$ (mg m$^{-3}$, SeaDAS4.8, OC4v4 algorithm) and in situ $C_{a}$ (mg m$^{-3}$). Grey circles and line: $C_{a}^{Fluo}$, blue diamonds and black solid line: $C_{a}^{HPLC}$. The dashed line shows the 1:1 relationship. The statistics of the comparisons are listed in Table 1.
Figure 2.4. Comparison between $C_a$ predicted by the OC4v4 algorithm (using SeaWiFS-derived $R_{rs}$ as input) and measured in situ $C_a$ (mg m$^{-3}$). Black broken line: OC4v4 prediction ($C_a^{SWF}$); grey circles and solid line: $C_a^{Fluor}$, blue diamonds and thick line: $C_a^{HPLC}$. 

$C_a^{Fluor}$: $n = 307$, $y = 4.83x^{-2.1}$, $R^2 = 0.71$.

$C_a^{HPLC}$: $n = 96$, $y = 2.05x^{-1.89}$, $R^2 = 0.45$. 

$Max(R_{rs443}, R_{rs490}, R_{rs510})/R_{rs555}$
Table 2.2. Statistics for the comparisons between $C_{\text{Fluor}}$ and $C_{\text{HPLC}}$ (mg m$^{-3}$) for data shown in Figure 2.5. $a_0$ and $a_1$ are the power fitting coefficients in the form of $C_{\text{Fluor}} = a_0 \times (C_{\text{HPLC}})^{a_1}$, $R^2$ is the corresponding coefficient of determination, $n$ is the number of matching pairs, RMS is root mean square error, SD is standard deviation.

<table>
<thead>
<tr>
<th>$C_{\text{HPLC}}$ range</th>
<th>0.01 - 15</th>
<th>&lt; 0.05</th>
<th>0.05 – 1.5</th>
<th>1.5 – 3.0</th>
<th>&gt; 3.0</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>832</td>
<td>21</td>
<td>647</td>
<td>96</td>
<td>68</td>
</tr>
<tr>
<td>$a_0$, $a_1$</td>
<td>1.40, 0.66</td>
<td>0.28, 0.14</td>
<td>1.34, 0.63</td>
<td>1.01, 0.96</td>
<td>2.15, 0.55</td>
</tr>
<tr>
<td>$R^2$</td>
<td>0.67</td>
<td>0.01</td>
<td>0.49</td>
<td>0.11</td>
<td>0.14</td>
</tr>
<tr>
<td>$C_{\text{Fluor}}/C_{\text{HPLC}} \pm$ SD</td>
<td>2.43 ± 3.37</td>
<td>10.06 ± 15.21</td>
<td>2.48 ± 2.23</td>
<td>1.15 ± 0.73</td>
<td>1.37 ± 1.04</td>
</tr>
<tr>
<td>RMS</td>
<td>366%</td>
<td>1739%</td>
<td>268%</td>
<td>74%</td>
<td>110%</td>
</tr>
<tr>
<td>bias</td>
<td>143%</td>
<td>905%</td>
<td>148%</td>
<td>15%</td>
<td>37%</td>
</tr>
<tr>
<td>log,RMS</td>
<td>0.40</td>
<td>0.87</td>
<td>0.40</td>
<td>0.23</td>
<td>0.34</td>
</tr>
<tr>
<td>log bias</td>
<td>0.25</td>
<td>0.79</td>
<td>0.29</td>
<td>-0.01</td>
<td>0.02</td>
</tr>
</tbody>
</table>
Figure 2.5. Comparison between \( C_a^{\text{HPLC}} \) and \( C_a^{\text{Fluor}} \) (mg m\(^{-3}\)) between January and February 1998 - 2001 (n = 832). Grey squares: \( C_a < 0.05 \) mg m\(^{-3}\), cyan circles: \( C_a \) between 0.05 - 1.5 mg m\(^{-3}\), green triangles: \( C_a \) between 1.5 - 3 mg m\(^{-3}\), blue diamonds: \( C_a > 3 \) mg m\(^{-3}\). The dashed line shows the 1:1 relationship. Statistics for the comparison are listed in Table 2.2.

\[
C_a^{\text{Fluor}} = 1.34(C_a^{\text{HPLC}})^{0.63}, \quad R^2 = 0.49, \quad n = 647
\]

\[
C_a^{\text{Fluor}} = 1.01(C_a^{\text{HPLC}})^{0.96}, \quad R^2 = 0.11, \quad n = 96
\]
Figure 2.6. Normalized histogram of $C_a^{SWF}$ distributions (mg m$^{-3}$) in the Southern Ocean during austral summer. (a) For the study region (Fig. 1.1) bound by 75 – 60º S and 75 – 60º W; (b) for the entire Southern Ocean (south of 60ºS). The y-axis shows the percentage surface area. 91% and 96% of the surface waters for (a) and (b), respectively, fall within the range of 0.05 to 1.5 mg m$^{-3}$.

**DISCUSSION**

Although HPLC has been recommended as the most reliable method to determine $C_a$ (e.g. Trees et al., 1985), most cruise surveys still use the fluorometric method because it is faster, requires less technical expertise and is less expensive than HPLC. The $C_a$ data originally used in the development of the OC4v4 algorithm (O’Reilly et al., 2000) included 2,853 in situ measurements from a variety of oceanic environments (but not the Southern Ocean), of which 72% were fluorometric and 28% were HPLC measurements. Therefore, the predicted $C_a$ satellite measurements should naturally lean toward the fluorometric values. However, this was not observed in the present study, suggesting that the species composition and their associated pigment absorption characteristics in waters west of the Antarctic Peninsula region may be different from the “mean” composition and absorption on which the original algorithm was based.
The large difference observed between $C_a^{\text{Fluor}}$ and $C_a^{\text{HPLC}}$ from the same water samples was likely due, in part, to interference of the fluorescence signal by chlorophyll accessory pigments ($C_b$, $C_c$ and their degradation products). In this study, $C_b$ only occurred in low concentrations compared to $C_a$ (mean ratio $C_b/C_a = 0.023$, $n = 486$); however, $C_c$ was relatively high (mean ratio $C_c/C_a = 0.25$, $n = 486$) (Fig. 2.7). The presence of significant amounts of $C_c$ is known to cause an overestimation of $C_a$ by the fluorometric method (Gibbs, 1979; Lorenzen, 1981).

$C_b$ is an accessory pigment in prochlorophytes, chlorophytes and prasinophytes, while $C_c$ is generally present in diatoms, dinoflagellates, cryptophytes and haptophytes (Parsons et al., 1984). Diatoms are the dominant phytoplankton in waters west of the Antarctic Peninsula, with dinoflagellates being very abundant at times (Prézelin et al, 2000, 2004). Prochlorophytes, a type of cyanobacteria first identified in the late 1980s (Chisholm et al., 1988), have not yet been observed in the Southern Ocean, while chlorophytes can be abundant (Prézelin et al, 2000, 2004). Similarly, cryptophytes are usually scarce in the water column, but can be very abundant in coastal surface melt water during spring and summer (Moline and Prézelin, 1996). Alloxanthin, the biomarker pigment for cryptophytes (Prézelin et al, 2000), occurred in 91% ($n = 516$) of the pigment samples. Hence, chlorophytes were probably the dominant source of $C_b$ during our study period, while the dominant sources of $C_c$ appear to be diatoms, dinoflagellates and cryptophytes, identified by the presence of fucoxanthin, peridinin, and alloxanthin in 99.5%, 53% and 91% of the samples, respectively.

$C_b$ and $C_c$ vary widely throughout the world’s ocean (Jeffrey, 1976; Lorenzen, 1981; Trees et al., 1985; Bidigare et al. 1986; Goericke and Repeta, 1993; Bianchi et al.,
1995). Overall, these studies found that $C_b$ can cause an underestimation of $C_a$ by the fluorometric method with ratios of $C_b/C_a$ ranging from 0.15 to 0.51, while the presence of significant amount of $C_c$ can lead to an overestimation of $C_a$. Typical ratios of $C_c/C_a$ for assemblages dominated by phytoplankton containing chlorophyll $c$ range from 0.15 to 0.44 (Bidigare et al., 1986; Bianchi et al., 1995; Lohrenz et al., 2003). Results reported here are consistent with these previous findings.

Can the presence of significant amount of $C_c$ lead to overestimation of $C_a$ when the latter is derived from remote sensing reflectance data? The inversion of remote sensing reflectance to $C_a$ is an implicit (e.g., OC4v4) or explicit (e.g., Maritorena et al., 2002) function of phytoplankton pigment absorption. Lohrenz et al. (2003) reported that even if the amount of accessory pigments (sum of carotenoids and $C_b + C_c$) is equal to $C_a$, the perturbation to the pigment absorption is < 30%, suggesting a relatively small error in the satellite-retrieved $C_a$. Hence, the large differences between $C_a^{\text{SWF}}$ and $C_a^{\text{Fluor}}$ observed here cannot be explained by the additional absorption of accessory pigment, but can be explained by the interference of these accessory pigments to the fluorescence peak when $C_a$ is determined using the fluorometric method.
Figure 2.7. Relationship between HPLC $C_b/C_a$ and $C_a^{\text{Fluor}}/C_a^{\text{HPLC}}$ ($y = 4.36 x^{0.26}$, $R^2 = 0.11$, $n = 482$), and between HPLC $C_c/C_a$ and $C_a^{\text{Fluor}}/C_a^{\text{HPLC}}$ ($y = 3.09 x^{0.39}$, $R^2 = 0.19$, $n = 482$). Note that the slope for the latter (0.39) is significantly larger than for the former (0.26). Here $C_b/C_a = 0.023 \pm 0.034$ ($n = 482$) and $C_c/C_a = 0.25 \pm 0.59$ ($n = 482$).

In summary, contrary to previous reports that estimates of $C_a^{\text{SWF}}$ in the Southern Ocean were significantly lower than those measured in situ, satellite estimates reported here agree with those determined from water samples for $C_a$ between 0.05 and 1.5 mg m$^{-3}$ for January-February between 1998 and 2001. This is primarily because the in situ $C_a$ data were determined by HPLC ($C_a^{\text{HPLC}}$) rather than by fluorometric methods ($C_a^{\text{Fluor}}$), which are known to introduce significant errors in $C_a$ estimates in the presence of certain accessory pigments.
CHAPTER THREE

SPATIAL AND TEMPORAL VARIABILITY OF SEAWIFS CHLOROPHYLL A DISTRIBUTIONS WEST OF THE ANTARCTIC PENINSULA: IMPLICATIONS FOR KRILL PRODUCTION

INTRODUCTION

Chlorophyll distributions in the Southern Ocean show high spatial and temporal variability. Most Antarctic open waters have relatively low chlorophyll concentrations despite the availability of nutrients and, thus, the Southern Ocean is generally considered to be a High-Nutrient Low-Chlorophyll region (Holm-Hansen et al., 1977). Nevertheless, large phytoplankton blooms do occur during spring and summer, particularly in waters associated with ice edges, polynyas, islands, and continental shelves (e.g., Smith and Nelson, 1985; Sullivan et al., 1993; Moore and Abbott, 2000; Arrigo and van Dijken, 2003). Some of the largest blooms develop in the marginal ice zone (MIZ), in waters associated with the seasonal advance and retreat of sea ice. As the ice edge recedes, low salinity meltwater produces a low density surface lens that reduces vertical mixing and shallows the mixed layer; consequently, phytoplankton are able to grow in a high irradiance stable environment. This phenomenon is common in the MIZ of the Weddell and Ross Seas, as well as in other regions around Antarctica (El-Sayed, 1971; El-Sayed and Taguchi, 1981; Smith and Nelson, 1985; Sedwick and DiTullio, 1997), with reported chlorophyll concentrations up to 190 mg m\(^{-3}\) (El-Sayed, 1971).
Polynyas are areas of open water surrounded by sea ice, usually characterized by elevated surface chlorophyll concentrations. Arrigo and van Dijken (2003) examined surface chlorophyll concentrations in 37 Antarctic polynyas and report values ranging from 0.24 to 7 mg chl m$^{-3}$ during summer. Maximum chlorophyll concentrations occurred in the Amundsen and Ross Seas, in agreement with previous reports of up to 40 mg m$^{-3}$ in the Ross Sea polynya (Arrigo and McClain, 1994). Antarctic islands can also be surrounded by chlorophyll-rich waters. For example, the region around South Georgia has been characterized as highly productive, with high densities of phytoplankton and zooplankton, as well as large colonies of seals and seabirds (Atkinson et al., 2001). Phytoplankton blooms in this area have been observed from November to April with chlorophyll concentrations reaching 30 mg m$^{-3}$. The high productivity at South Georgia has been attributed to a combination of factors including enhanced supply of iron and rapid recycling of nitrogen, favorable temperatures, and a shallow stable inshore water column (reviewed in Atkinson et al., 2001).

The continental shelf waters west of the Antarctic Peninsula (55 - 75°S, 50 - 80°W) (Fig. 3.1) in particular, are considered to be one of the most productive regions of the Southern Ocean, supporting high densities of phytoplankton, zooplankton, and upper trophic level predators (e.g., Fraser and Trivelpiece, 1996; Arrigo et al. 1998; Deibel and Daly, 2007). Chlorophyll concentrations in the northern sectors along the Antarctic Peninsula shelf have been reported to reach values up to 38 mg chl m$^{-3}$ during December and January (Holm-Hansen and Mitchell, 1991; reviewed in Smith et al., 1996a; Rodriguez et al., 2002). More recently, Arrigo and van Dijken (2003) reported that SeaWiFS chlorophyll in a phytoplankton bloom related to the Marguerite Bay polynya
averaged 2.30 mg chl m\(^{-3}\), while Garibotti et al. (2003) and Meyer et al. (2003) observed summer chlorophyll concentrations up to 17.86 mg m\(^{-3}\) in 1997 and 25 mg m\(^{-3}\) in 2000, respectively, within Marguerite Bay. These findings suggest that the southern sector of the Antarctic Peninsula also may support large phytoplankton blooms.

Studies of the spatial and temporal dynamics of phytoplankton along the northern part of the Peninsula reveal a pattern of chlorophyll accumulation in coastal areas over the summer, with lower concentrations offshore (Smith et al., 1998a; Garibotti et al., 2003). These studies also identify an alongshore gradient with higher biomass in the northern sectors of the Antarctic Peninsula earlier in the productive season, which later progresses to the southeast as the sea ice retreats in the same direction. Most studies to date, however, have only dealt with chlorophyll distributions north of Marguerite Bay and, thus, information on phytoplankton dynamics in the vicinity of Marguerite Bay and to the south in the Bellingshausen Sea is scarce.

The Southern Ocean Global Ocean Ecosystems Dynamics (SO GLOBEC) program investigated the physical and biological factors that influence the growth, recruitment, and overwintering survival of Antarctic krill, *Euphausia superba*, in the vicinity of Marguerite Bay (Fig. 3.1), west of the Antarctic Peninsula, during austral fall and winter of 2001 and 2002. Krill play a key role in the Antarctic ecosystem as one of the primary pelagic herbivores and prey for many predators. The large krill population west of the Antarctic Peninsula appears to be maintained by occasional strong year classes, with often poor recruitment in the intervening years (Siegel and Loeb, 1995; Hewitt et al., 2003; Quetin and Ross, 2003). The suite of physical and biological factors that govern krill reproduction and recruitment, however, remain poorly known.
During the GLOBEC study, large differences in abundances of larval and juvenile krill were observed between the two years (Daly, 2004). During fall 2001, larvae were very abundant (< 0.01 - 132 ind m\(^{-3}\)), with younger stages dominant offshore and older stages dominant onshelf (Fig. 3.2, see Fig. 3.1 for station locations). Offshore larval densities (up to 132 ind m\(^{-3}\)) are amongst the highest reported for the area and are comparable to those reported by Rakusa-Suszczewski (1984) for 1981. Few juveniles were observed anywhere in 2001. During fall 2002, relatively high concentrations of young larvae were again detected in oceanic waters (< 0.01 - 211 ind m\(^{-3}\)), although average abundances were significantly lower than in 2001 and all larval stages were scarce in coastal areas (Fig. 3.2). In contrast, juveniles were relatively abundant on the middle and inner shelf in the vicinity of Marguerite Bay (< 0.01 - 2.37 ind. m\(^{-3}\)), indicating a successful recruitment from the 2001 larval population. These results led me to investigate the environmental conditions that contributed to the large krill reproduction during austral spring and summer 2000/2001, and subsequent high larval densities.

Herein, I investigate chlorophyll dynamics west of the Antarctic Peninsula using SeaWiFS ocean color data between 1997 and 2004, with special emphasis on the Marguerite Bay region, to better understand the conditions that make it a suitable habitat for krill. I also examine the effects of the retreat of the ice edge on the timing and location of phytoplankton blooms west of the Antarctic Peninsula. Finally, I discuss the environmental mechanisms that potentially support successful krill reproduction and recruitment in this area.
METHODS

The study area consisted of the coastal waters west of the Antarctic Peninsula, adjacent deep waters in the Drake Passage, and coastal and oceanic waters of the Bellingshausen Sea (55 - 75 °S and 50 - 80 °W), as chlorophyll concentrations in these areas are most likely to influence regional krill populations (Fig. 3.1).

Krill were collected aboard the R.V. Lawrence M. Gould during austral autumn between 23 April and 6 June 2001 and between 7 April and 20 May 2002 as part of the US SO GLOBEC program. A total of 18 and 16 net tows were done at several stations during 2001 and 2002 respectively. Individuals were collected at eight discrete depth intervals using a 1-m² MOCNESS (Multiple Opening-Closing Net and Environmental Sensing System) net system, with a 333 μm mesh. Krill were identified for stages of larvae (calyptopis I - III, furcilia I - VI), juveniles, or adults (males or females) after Makarov (1980) and measured for total length (from the base of the eye to the tip of the telson, excluding setae). Herein, the distribution of krill stage abundances are compared at two representative stations; one offshore (Sta. 1) and one onshelf (Sta. 5) (Fig. 3.2). Abundances are the mean of four onshelf and two offshore net tows in 2001, and three onshelf and two offshore net tows in 2002. Stage abundances (A, ind m⁻³) were calculated as a weighted mean for the sampling depth using:

\[ A = \frac{\sum_{i=1}^{8} x_i * z_i}{\sum_{i=1}^{8} z_i} \]

where \( i \) represents each of the eight nets (collections from different depth strata) in each
cast, \( x \) is the abundance of each krill stage in each net (ind m\(^{-3}\)), and \( z \) is the depth interval of each stratum (m).

A recruitment index (\( R_1 \)) for *E. superba* was calculated as the proportion of one year-old krill compared to age-class one plus all older age classes from all net samples for fall 2001 and 2002. Age-class one was defined as juvenile krill with total length ranging from 20 - 30 mm following Siegel et al. (1998). The total abundance (ind m\(^{-3}\)) of juveniles 20 - 30 mm and of juveniles 20 - 30 mm plus all older stages was estimated for each net and summed over the entire cruise to estimate a recruitment index for each cruise. Offshelf net tows (Sta. 1), which did not contain any juvenile or adult krill, were excluded from the recruitment calculation. All except one of the onshelf stations were located in inner coastal waters; therefore, recruitment estimates were not biased as a result of migration of large krill to the inner shelf in late summer and fall (Siegel et al., 2003; Hewitt et al., 2003).

The chlorophyll dataset includes 6606 SeaWiFS daily Level 2 files (~ 1 km/pixel near nadir) between September 1997 and December 2004 obtained from NASA Goddard Space Flight Center. These data were collected by ground stations, as well as occasional satellite onboard recording over the area using the most recent algorithms and software package (SeaDAS4.8). The level 2 data were mapped to a rectangular projection with approximately 1 km\(^2\)/pixel for the western Antarctic Peninsula region (Fig. 3.1). The parameter used in this study is the surface chlorophyll concentration derived from the OC4v4 empirical band-ratio (blue versus green) algorithm (O’Reilly et al., 2000). SeaWiFS chlorophyll estimates are biased toward surface values as the sensor only “sees” the first few meters of the water column. Nevertheless, SeaWiFS chlorophyll provides a
good assessment of water column concentrations in the Southern Ocean, as maximum chlorophyll concentrations are near surface and surface chlorophyll is well correlated with depth integrated chlorophyll (Holm-Hansen and Mitchell, 1991; Holm-Hansen et al., 2004a; Korb et al., 2004).

The accuracy of the SeaWiFS algorithm in the Southern Ocean is being debated. Several studies using \textit{in situ} chlorophyll fluorescence determined that the SeaWiFS algorithm underestimates chlorophyll $a$ concentrations in the Southern Ocean (e.g., Dierssen and Smith, 2000; Korb et al., 2004). However, recent results from more accurate \textit{in situ} HPLC (High Performance Liquid Chromatography) values demonstrated that SeaWiFS chlorophyll $a$ estimates are accurate (bias = 12\%) for chlorophyll concentrations between 0.1 and $\sim$ 4 mg m$^{-3}$, which include $>$ 90\% of the waters in the Southern Ocean (Chapter 2; Marrari et al., 2006). The mean ratio between SeaWiFS chlorophyll and \textit{in situ} HPLC chlorophyll was close to unity (i.e., 1.12) ($n = 96$), indicating good agreement between the two sets of data. Although these previous results could not verify the SeaWiFS algorithm for chlorophyll concentrations greater than $\sim$ 4 mg m$^{-3}$, here I use the algorithm to estimate a larger range of chlorophyll concentrations in order to investigate relative changes in chlorophyll spatial and temporal dynamics. In addition, because colored dissolved organic matter (CDOM) may introduce significant errors to the estimation of surface chlorophyll $a$ from ocean color remote sensing data (e.g., Carder et al., 1989), I measured CDOM absorption from 10 surface water samples collected in the study area between April 4 and May 10, 2002. Absorbance spectra were obtained at 1 nm intervals from 200 to 750 nm with a Hitachi U-3300 double-beam spectrophotometer with 10-cm quartz cells. MilliQ water was used in the reference cell. Three scans were run for
Figure 3.1. (a) Map of the Antarctic continent showing the location of the study area and other geographic references. The solid line represents the 1000 m isobath and the dashed line indicates the Antarctic Circle (66.3°S). (b) Details of the Antarctic Peninsula region and location of the US Southern Ocean GLOBEC offshelf (Sta. 1) and onshelf (Sta. 5) net sampling stations (●) represented in Fig. 2.
each sample and the resulting spectra were averaged to reduce noise. Data were corrected for scattering by subtracting absorbance at 700 nm from all measurements. Absorbance values were converted to absorption coefficients (Kirk, 1983). The absorption coefficient at 375 nm, $a_{375}$, was used as an index of CDOM concentration. Because $a_{375}$ was low in all samples ($0.02 - 0.148 \text{ m}^{-1}$), I conclude that the accuracy of the SeaWiFS chlorophyll estimates is not affected by the presence of significant levels of CDOM.

The Southern Ocean daily satellite images normally have missing data due to a relatively high percentage of cloud cover; therefore, at least weekly composites of data typically are needed to obtain good spatial coverage (e.g., Holm-Hansen et al., 2004a). In this study, biweekly composites of SeaWiFS chlorophyll data are used. A seven-year climatology of biweekly chlorophyll $a$ concentrations (mean chlorophyll concentration at each pixel) was generated from the mapped Level 2 data from September 1997 through December 2004 and biweekly composite images were produced. When calculating the mean value in either the climatology or the biweekly data, only valid data were used. Suspicious data identified by various quality flags associated with each pixel (for example cloud contamination, large solar/view angle, etc.) were excluded from the calculations.

The study area was divided into fourteen subregions, each representing different oceanographic conditions in order to (1) investigate the initiation and progression of phytoplankton blooms, and (2) analyze the relative differences in chlorophyll concentrations between regions along the coastal Antarctic Peninsula and offshore areas. Regions 1 - 6 represent offshelf oceanic regimes with depths greater than 2000 m, while regions 7 and 8 are located over the continental shelf slope, defined as the area between 500 and 2000 m. Regions 9, 10 and 11 represent coastal waters along the Antarctic
Peninsula shelf and regions 13 and 14 are located in Marguerite Bay. Region 12 includes both coastal and oceanic waters in the Scotia-Weddell confluence area. The biweekly geometric mean chlorophyll concentrations between September - March 1997 to 2004 were calculated for each region and the results plotted over time in relation to the climatology. Although values up to 55 mg m\(^{-3}\) were estimated from SeaWiFS for Marguerite Bay during summer, concentrations lower than 0.01 mg m\(^{-3}\) and greater than 20 mg m\(^{-3}\) were excluded from the geometric mean calculations, since the accuracy of these values could not be verified with concurrent \textit{in situ} data. However, as most of the SeaWiFS chlorophyll concentrations during the study period were within the 0.01 - 20 mg m\(^{-3}\) range, the geometric mean calculations were not significantly affected by excluding the few extreme values. Gaps in the time series occurred when the available data points were < 10% of the total number of pixels within each region during the biweekly period of maximum spatial coverage.

The mean location of the ice edge within the study area during October, November, and December of 2000 and 2001 was determined using a two-dimensional linear interpolation of monthly ice concentration on a 25 km resolution grid. Monthly averaged gridded ice concentrations generated using the NASA Team algorithm and Nimbus-7 SMMR and DMSP SSM/I passive microwave data were obtained from the National Snow and Ice Data Center (Cavalieri et al., 2005). The ice edge was considered to be the location where sea ice concentration was \(\leq 15\%\) (Gloersen et al., 1992). The mean location of the ice edge during these months was superimposed over the concurrent biweekly SeaWiFS chlorophyll images. In addition, the mean location of the ice edge during the preceding month also was plotted in order to evaluate changes in chlorophyll
concentrations within the region of ice edge retreat. To examine whether there was a relationship between the retreat of the ice edge and the formation of phytoplankton blooms, chlorophyll concentrations along the location of the ice edge in September, October, and November 2000 and 2001 were extracted from the satellite data. The chlorophyll values at the same locations were extracted for the subsequent 6-biweekly periods (three months). Thus, at the September location of the ice edge, chlorophyll concentrations were analyzed during September - November and at the November ice edge location, chlorophyll was analyzed during December - January. The median chlorophyll concentration for each biweekly period was estimated and plotted over time to generate a time series of chlorophyll concentrations at the ice edge location.

The daily area free of sea ice (km²) in the northern and southern sections of Marguerite Bay was estimated from satellite data following the methods described in Arrigo and van Dijken (2003). A daily climatology from 1997 through 2004 was calculated as the mean daily ice-free area within each subregion. A running average was applied to reduce the daily variability (bandwidth = 9.1 days). The time series of daily ice-free area from 1997 to 2004 were plotted in relation to the 8-year climatology with special emphasis on 2001 and 2002.
Figure 3.2. Mean abundance (ind m\(^{-3}\)) of development stages of *Euphausia superba* in offshelf waters (Sta. 1) west of the Antarctic Peninsula and coastal waters (Sta. 5) of Marguerite Bay during fall 2001 (light grey) and 2002 (dark grey). C1 - CIII = calyptopis 1 - 3; F1 - FVI = furcilia 1 - 6; juvs = juveniles; males = adult males; females = adult females.
RESULTS

Biweekly climatological patterns of chlorophyll $a$ indicate that oceanic and coastal areas in the Bellingshausen Sea and coastal Marguerite Bay had persistently high chlorophyll concentrations ($0.1 - > 7$ mg m$^{-3}$) during spring and summer in comparison with any other area west of the Antarctic Peninsula between September and March 1997 - 2004 (Fig. 3.3). Oceanic waters offshore of the northern Antarctic Peninsula typically had relatively low concentrations ($0.1 - 0.2$ mg m$^{-3}$). Intermediate values ($0.1 - ~ 2$ mg m$^{-3}$) were generally observed over the more northern continental shelf regions west of the Antarctic Peninsula and downstream in the Scotia Sea, although small, short-lived blooms with chlorophyll values greater than 2 mg m$^{-3}$ occurred nearshore in this region.

The spatial and temporal changes in chlorophyll patterns suggest that biomass accumulations initially occurred during October and November in offshelf waters, mainly in the Bellingshausen Sea and to a lesser extent near the shelf break in the vicinity of the Shetland Islands at the northern end of the Antarctic Peninsula (Fig. 3.3). As the season progressed (mid-December), phytoplankton blooms developed onshore, especially in the vicinity of Marguerite Bay and the coastal Bellingshausen Sea, where they remained well established until early April. In contrast, there didn’t appear to be a significant seasonal increase in chlorophyll in northern Peninsula waters, except near some of the islands such as the South Shetlands and a few short-lived blooms nearshore.

A cross-shelf gradient in chlorophyll concentrations (higher values in coastal areas to lower values at the shelf break) was observed for Marguerite Bay and to the south during summer (January - March). Between Anvers and Adelaide islands, there was a similar cross-shelf gradient in January, but by late February maximum chlorophyll
concentrations had shifted to mid-shelf and outer shelf areas near Anvers Island. At the northern end of the Peninsula, there was no apparent cross-shelf gradient. Instead, maximum chlorophyll values occurred in the vicinity of the Shetland Islands and Elephant Island near the outer shelf.

The geometric mean chlorophyll concentrations for the 14 subregions defined in the study area (Fig. 3.4a) illustrate the interannual (1997 - 2004) variability in patterns of chlorophyll distribution west of the Antarctic Peninsula. Variations from the climatology were relatively small (maximum ± 0.4 mg m⁻³) in offshelf regions of the Antarctic Circumpolar Current (ACC), Drake Passage, and the Scotia Sea (regions 1 - 6), with the Bellingshausen and Scotia Sea regions having higher variability than the Drake Passage regions (Fig. 3.4b). Regions 1 - 2 and 4 - 5 had a small chlorophyll peak in early November (year day 315), whereas the Scotia Sea generally sustained higher levels of chlorophyll for longer periods (until early April). Variability in waters at the tip of the Peninsula (region 12) was influenced by the large chlorophyll concentrations to the east of the Antarctic Peninsula in the Weddell Sea. Shelf slope waters (regions 7 and 8) had similar interannual ranges in chlorophyll, although the years with chlorophyll concentrations greater than climatology were not necessarily the same. For waters over the continental shelf (regions 9 - 11), the southern Bellingshausen and Marguerite Bay shelf region had the highest chlorophyll concentrations and variability, with declining values to the northeast.
Figure 3.3. Biweekly climatology (1997 - 2004) of SeaWiFS chlorophyll concentrations (mg m$^{-3}$) between October and March. The date range included in each image is indicated on the lower right hand corner. White areas indicate no data due to the presence of clouds and/or sea ice. The white thin line represents the 1000 m isobath.
The highest chlorophyll concentrations at the northern tip of the Peninsula (regions 3, 6, and 12) occurred during 1999/2000. Elsewhere, 2000/2001 had substantially higher chlorophyll concentrations during summer (December - February) compared with most of the other years analyzed, particularly in the Bellingshausen Sea (regions 1, 4, 7 and 9) and Marguerite Bay (regions 13 and 14). In waters over the continental shelf of the Bellingshausen Sea (regions 7 and 9), chlorophyll was generally elevated during the spring and summer seasons of both 2000/2001 and 2001/2002, with high variability observed both between and within years. During 2001/2002, however, these chlorophyll peaks did not occur for as extended a period of time as in 2000/2001. Shelf break and
coastal regions along the Antarctic Peninsula (regions 8, 10 and 11) had elevated mean chlorophyll concentrations relative to offshore areas, although the variations (up to 0.41 mg m$^{-3}$) with respect to the climatology were less evident than in the Bellingshausen Sea (region 9, up to 0.97 mg m$^{-3}$) or Marguerite Bay (up to 3.72 mg m$^{-3}$), and mean values never exceeded 0.72 mg m$^{-3}$ (note the different y-axis scales between plots). Marguerite Bay had the highest chlorophyll concentrations in comparison with any other region analyzed. In northern Marguerite Bay, average values during January for all years were 1.24 - 1.31 mg m$^{-3}$. However, during January 2001, mean chlorophyll reached 4.95 mg m$^{-3}$, whereas 2002 values were consistently near or below the 7-year average. In southern Marguerite Bay, 2000/2001 showed high mean values of up to 2.76 mg chl m$^{-3}$ from late December through February, a factor of 2.5 higher than average (1.09 mg chl m$^{-3}$) conditions. The presence of sea ice prevented satellite chlorophyll data collection during most of the 2001/2002 summer.

The location, timing, and extent of sea ice during 2000/2001 and 2001/2002 were examined in relation to chlorophyll concentrations to better understand the relationship between sea ice and phytoplankton blooms (Fig. 3.5). Chlorophyll concentrations were highly variable in relation to the receding ice edge in our study area. During September 2000 and 2001, the ice edge was located in oceanic waters of the ACC. By October, the ice margin had retreated considerably and occurred closer to the coast at its eastern extent, but chlorophyll had not increased significantly at the September ice edge locations (Fig. 3.5a and 3.5b, top 2 panels). This suggests that October was too early in the productive season for any significant chlorophyll accumulations to occur within the ice edge zone. In November, the ice edge had receded onshelf in the mid-Antarctic Peninsula, but in the
Bellingshausen Sea the ice edge remained offshore in 2000 and approximately at the shelf break in 2001. Chlorophyll concentrations increased significantly in this region, reaching ~ 5 mg m\(^{-3}\) (Fig. 3.5a and 3.5b, center panels). Although ice edge blooms appear to have occurred in some parts of the Bellingshausen Sea during November and December, most of this region of enhanced chlorophyll was presumably too far from the ice edge to have been influenced by ice melt processes.

Figure 3.4a. Location of the 14 subregions along the western Antarctic Peninsula superimposed over the climatology (1998 - 2004) of SeaWiFS chlorophyll concentrations (mg m\(^{-3}\)) for January 1 - 14.
Figure 3.4b. Time series of geometric mean chlorophyll concentrations in each subregion for each biweekly period during 1997 - 2004. The 7-year climatology is also included for each region (thick black line with circles). Note the difference in scale of the y-axis for the different regions. The 2000/2001 (red squares), 2001/2002 (blue circles), 2002/2003 (cyan broken line), and 2003/2004 (green dotted line) spring-summer seasons are highlighted in color.
Figure 3.5a. Biweekly SeaWiFS chlorophyll concentrations (Chl, mg m\(^{-3}\)) in October (Oct), November (Nov) and December (Dec) of 2000. The mean monthly location of the ice edge is also shown: the red line indicates the location of the ice edge during the preceding month, the yellow line represents the current month. The 1000 m isobath is indicated by the white line.
Figure 3.5b. Biweekly SeaWiFS chlorophyll concentrations (Chl, mg m$^{-3}$) in October (Oct), November (Nov) and December (Dec) of 2001. The mean monthly location of the ice edge is also shown: the red line indicates the location of the ice edge during the preceding month, the yellow line represents the current month. The 1000 m isobath is indicated by the white line.
The analysis of chlorophyll buildup during the weeks following the retreat of the ice edge reveals an increase in chlorophyll concentrations in the vicinity of the October and November ice edge locations, both in 2000 and 2001; however, values usually reached a maximum 4 - 6 weeks after the ice had receded (Fig. 3.6). In addition, other areas along the Antarctic Peninsula that were never influenced by sea ice also showed a similar increase during our study period. For example in November and December of 2001 (Fig. 3.5b), the ice edge occupied coastal areas of the Bellingshausen Sea and along the Antarctic Peninsula to Anvers Island (see Fig. 3.1 for site locations). Even though ice never occupied the northern end of the Peninsula, elevated chlorophyll concentrations were observed along the shelf break and in coastal areas. Thus, processes other than the retreat of the ice edge likely influenced phytoplankton dynamics in this more northern area.

Figure 3.6. Biweekly time series of chlorophyll accumulation after the ice had receded at the September, October, and November 2000 and 2001 locations of the ice edge shown in figure 3.5.
Sea ice coverage in Marguerite Bay also showed strong variability between the years analyzed and these differences are particularly marked between 2000/2001 and 2001/2002. During summer and early fall (January - April), typical values of ice-free areas range from approximately 9,000 to 11,000 km² in northern Marguerite Bay and from ~ 4,500 to 7,500 km² in southern Marguerite Bay (Fig. 3.7c). A comparison of the climatology and 2001 and 2002 daily ice-free areas (km²) indicates that 2002 had above average sea ice in both the northern and southern sectors throughout the spring, summer, and fall (Fig. 3.7a and 3.7b). In addition, ice formed earlier in 2002 than in 2001. In contrast, 2001 had sea ice values significantly below the 8-year mean, particularly from January through July, as indicated by the unusually large ice-free areas observed both in the northern and southern sectors. In 2001 these values reached approximately 12,000 and 11,500 km² in the northern and southern regions respectively. On the other hand, the areas free of ice only reached 6,000 - 9,000 km² in the northern and 0 - 2,000 km² in southern sectors during the same months in 2002, suggesting an especially extensive sea ice cover. During winter (starting in mid-July or year day ~ 200), sea ice conditions were similar for both years, although, as mentioned above, ice occupied both the northern and southern sectors considerably earlier in 2002. Other years with above-average sea ice cover were 1997/1998 and 1999/2000, while during 1998/1999 and 2003/2004 sea ice was lower than normal.

Recruitment indices (R₁) for *E. superba* collected during fall in the GLOBEC study area were 0 for 2001 and 0.4 for 2002, representing no juvenile recruitment from the 1999/2000 larval year class and significant recruitment from the 2000/2001 larval year class, respectively. Published krill recruitment index values for years of elevated
recruitment west of the Antarctic Peninsula between 1975 and 2002 are provided for comparison (Table 3.1).

Figure 3.7. Daily ice-free area (km$^2$) in (a) northern and (b) southern Marguerite Bay during 1997 - 2004. The 8-year climatology also is shown (black thick line). (c) Location of the northern (dark grey) and southern (light grey) Marguerite Bay regions.

Figure 3.7. Daily ice-free area (km$^2$) in (a) northern and (b) southern Marguerite Bay during 1997 - 2004. The 8-year climatology also is shown (black thick line). (c) Location of the northern (dark grey) and southern (light grey) Marguerite Bay regions.
Table 3.1. Years of elevated krill recruitment between 1975 and 2002. Source publications are designated by symbols in the table. Recruitment indices ($R_1$) were estimated from the proportion of one year-old krill compared to age-class one plus all older age classes. Year class is the year larvae were produced; whereas recruitment to the juvenile stage occurs the following year. All krill were collected during austral summer (January - March) in waters west of the Antarctic Peninsula, except this study when krill were collected during austral fall (April - June). Only years of high recruitment ($R_1 > 0.5$) are shown.

<table>
<thead>
<tr>
<th>Year class</th>
<th>$R_1$</th>
<th>Location</th>
<th>Study period</th>
</tr>
</thead>
<tbody>
<tr>
<td>1979/1980</td>
<td>0.559</td>
<td>Elephant Island area *</td>
<td>1975 - 1996</td>
</tr>
<tr>
<td>1980/1981</td>
<td>0.757</td>
<td>Elephant Island area *</td>
<td>1975 - 1996</td>
</tr>
<tr>
<td>1987/1988</td>
<td>0.651</td>
<td>Elephant Island area *</td>
<td>1977 - 1997</td>
</tr>
<tr>
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<td>0.622</td>
<td>Elephant Island area *</td>
<td>1975 - 1996</td>
</tr>
<tr>
<td></td>
<td>0.639</td>
<td>Western Antarctic Peninsula #</td>
<td>1985 - 2002</td>
</tr>
<tr>
<td>1999/2000</td>
<td>0.573</td>
<td>Elephant Island area *</td>
<td>1975 - 2000</td>
</tr>
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<td></td>
<td>0</td>
<td>Western Antarctic Peninsula #</td>
<td>1985 - 2002</td>
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<tr>
<td></td>
<td>0</td>
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<td>2001 - 2002</td>
</tr>
<tr>
<td>2000/2001</td>
<td>0.748</td>
<td>Western Antarctic Peninsula #</td>
<td>1985 - 2002</td>
</tr>
<tr>
<td></td>
<td>0.400</td>
<td>Marguerite Bay region §</td>
<td>2001 - 2002</td>
</tr>
</tbody>
</table>

* Siegel et al. (2002); # Siegel et al. (2003); § this study
DISCUSSION

Chlorophyll concentrations showed considerable temporal and spatial variability in waters west of the Antarctic Peninsula during austral spring and summer between 1997 and 2004, with the largest and most persistent phytoplankton blooms consistently occurring in Marguerite Bay and the Bellingshausen Sea areas. The climatology showed a cross-shelf chlorophyll gradient in early summer in the middle and southern regions of the Peninsula. Investigators from the Palmer Long Term Ecological Research (LTER) program have reported a similar gradient in chlorophyll concentrations for January 1991-1995 (Smith et al., 1998a) and January 1997 (Garibotti et al., 2003) between Anvers and Adelaide islands, suggesting that this pattern of chlorophyll distribution has remained relatively constant in this area for about two decades. In 1997, average chlorophyll concentrations in coastal areas were an order of magnitude greater than those in the vicinity of the shelf break (4.38 mg m\(^{-3}\) vs. 0.22 mg m\(^{-3}\)) and corresponded to a shallower mixed layer and greater vertical stability of the water column in coastal waters (Garibotti et al., 2003). Climatology results presented here indicate that these environmental conditions may usually change by mid-February in the middle region of the Peninsula (Anvers to Adelaide islands) as this cross-shelf pattern was no longer present, except in Marguerite Bay and to the south.

The above mentioned studies also describe an alongshore gradient in chlorophyll concentrations with higher values in the northern sectors of the Peninsula earlier in the season possibly associated to the seasonal alongshore retreat of the sea ice and/or to latitudinal differences between areas. Present results, however, indicate that chlorophyll concentrations in the southern sectors are consistently higher than in any other area.
analyzed west of the Antarctic Peninsula. In addition, chlorophyll accumulations in these areas occur earlier in the spring and persist longer throughout the summer than most areas in the northern regions. Thus, the southern areas are vitally important to the Antarctic Peninsula ecosystem in terms of overall chlorophyll standing stock and the phytoplankton blooms are likely to play an important role in supporting higher trophic level dynamics.

Few studies provide information on phytoplankton dynamics in the Bellingshausen Sea (Savidge et al., 1995; Barlow et al., 1998). SeaWiFS images of the circumpolar distribution of mean annual chlorophyll concentrations show that the Bellingshausen and Amundsen Seas support large phytoplankton blooms (Fig. 1 in El-Sayed, 2005). Satellite derived estimates of primary productivity also indicate that the Bellingshausen/Amundsen Sea area is one of the most productive in Antarctic waters, only exceeded by the Ross and Weddell Seas (Arrigo et al., 1998). Typically, ice edge blooms are tightly coupled to spring ice edge retreat in Antarctic waters (Sullivan et al., 1993; Arrigo and McClain, 1994; Garibotti et al., 2005), where blooms develop within about two weeks after the ice recedes from a particular location. During austral spring 1992, investigators from the UK STERNA Program observed elevated chlorophyll concentrations in oceanic waters of the Bellingshausen Sea in late November (up to > 7 mg m⁻³) and early December (up to 2.4 mg m⁻³), but reported that they were not related to sea ice retreat (Savidge et al., 1995; Barlow et al., 1998). During the present study, some blooms appeared to occur along the ice edge in the Bellingshausen Sea; however, most blooms occurred with approximately a 4 - 6 week lag and, therefore, probably were not related to the ice retreat, in agreement with the STERNA Program observations. Clearly, there is a need for future phytoplankton studies in the Bellingshausen Sea in order to elucidate the factors that
control the formation and persistence of spring blooms in the region.

Ice edge blooms have been suggested to be an important feature in the northern Antarctic Peninsula region (e.g., Siegel and Loeb, 1995; Smith et al., 1998b). In contrast, findings reported herein indicate that the formation of spring blooms was not necessarily coupled to the retreat of the ice edge in the vicinity of the northern Peninsula between 1997 - 2004. Instead, blooms first appeared near the shelf break and gradually progressed to more coastal areas, suggesting that shelf break processes were likely an important factor influencing phytoplankton growth in the northern Peninsula region. The strong currents of the eastward flowing ACC interact with the bathymetry when it encounters the shelf break, generating meanders that usually can be detected at the surface. Antarctic surface waters are rich in macronutrients; however, iron deficiency has been proposed as a factor limiting phytoplankton growth (De Baar et al., 1995; Holm-Hansen et al., 2004b; 2005). The importance of upwelled iron-rich deep ACC waters to chlorophyll aggregations has been described for several regions of the Southern Ocean, including the Scotia Sea, the Polar Front region downstream of South Georgia, the Ross Sea, and the Antarctic Peninsula shelf break (De Baar et al., 1995; Measures and Vink, 2001; Prézelin et al., 2000; 2004; Holm-Hansen et al., 2005). Hence, upwelling of iron-rich deep water, rather than the retreat of the ice edge, may be a major factor controlling phytoplankton bloom development during spring and summer in the vicinity of the shelf break, and in coastal waters along the northern Antarctic Peninsula.

Although sea ice extent and duration in the Bellingshausen Sea and along the Antarctic Peninsula has decreased over the past 25 years (Parkinson, 2002; Ducklow et al., 2006), high interannual variability in sea ice is still observed. Ducklow et al. (2006)
analyzed 14 years (1991 - 2004) of sea ice extent data near Palmer Station in the vicinity of Anvers Island, and found that 2001 had the lowest (69,932 km²) winter sea ice extent of all years analyzed, while 2002 had the highest (109,936 km²) (mean = 91,112 km²). The early retreat of sea ice in Marguerite Bay in spring of 2001 resulted in large phytoplankton blooms in ice-free waters during summer. In contrast, the persistent presence of sea ice in Marguerite Bay during summer - fall 2002 resulted in overall lower chlorophyll concentrations in coastal surface waters. The positive relationship between sea ice cover and chlorophyll concentrations is further supported by observations during other years of this study. For example in 2003/2004, peaks of above-average chlorophyll concentration in northern and southern Marguerite Bay during January (Fig. 4b; regions 13 and 14) coincided with an early retreat of the sea ice in late 2003 and lower than normal sea ice extent during late 2003 - early 2004, particularly in the northern region (Fig. 3.7). In addition, the ice-free area in northern Marguerite Bay during late 1998 - early 1999 was larger than average, concurrent with high chlorophyll concentrations in the region during December - January 1998/1999.

Several investigations have suggested that sea ice extent and duration are the primary environmental factors influencing krill recruitment in the northern regions of the Antarctic Peninsula, as spring - summer ice edge blooms were believed to support krill reproduction and winter sea ice biota to provide food for overwintering larvae (Kawaguchi and Satake, 1994; Siegel and Loeb, 1995; Quetin and Ross, 2003). Results from this study, however, indicate that ice edge blooms are not prevalent in this region and, thus, may not be a primary source of food for reproducing krill. In addition during both winters of the GLOBEC study, sea ice biota concentrations were very low (0.05 -
0.07 mg chl m$^{-3}$) at the ice-water interface where larval krill feed (Daly, 2004). Indeed, a large percentage of larvae were not even associated with the undersurface of sea ice and instead remained in the water column, particularly in 2002. Also in both winters, larvae showed evidence of food limitation, as indicated by delayed development, decrease in growth rates, increased intermolt period, and decrease in dry weight, body carbon, and nitrogen (Daly, 2004). Hence, the presence of extensive sea ice during winter is not necessarily a good predictor of food availability for overwintering larvae and, environmental factors other than winter sea ice conditions must play an important role in recruitment.

Krill typically reproduce during late spring and summer (November - March) west of the Antarctic Peninsula (Siegel, 1988). Results from net samples suggest that the majority of the females migrate near the vicinity of the shelf break where they spawn in oceanic waters (e.g., Siegel, 1992; Hoffman et al., 1992), possibly owing to the predictable shelf break blooms that occur early in the productive season as observed in the climatology (Fig. 3.3). Successful krill reproduction and larval survival require an adequate food supply (Ross and Quetin, 1983; 1989). Adult females may require above average phytoplankton concentrations (1 - 5 mg chl m$^{-3}$) to initiate reproduction (Ross and Quetin, 1986) and relatively high chlorophyll concentrations (> 0.5 mg chl m$^{-3}$) to sustain multiple spawning throughout the summer (Nicol et al., 1995). It also is critical for the first-feeding larvae (calyptopis I) to encounter an adequate food supply in the euphotic zone within 10 - 14 days, otherwise they will not survive (Ross and Quetin, 1986). Hence, knowledge about differences in the timing, extent, and evolution of phytoplankton blooms is critical for understanding the interannual variability observed in krill.
recruitment success.

The recruitment indices in Table 1 indicate that only six out of 27 years had successful recruitment. Our fall (April - May) estimated recruitment ($R_1 = 0.4$) for 2000/2001 is lower than the 0.748 reported for summer by Siegel et al. (2003) (Table 3.1). A decrease in krill recruitment indices is commonly observed between summer and fall (Siegel and Loeb, 1995), owing to the seasonal decline in krill population abundance (Lascara et al. 1999). The lower fall indices correlate with summer values and, therefore, can still serve as a relative indicator of recruitment. The two highest recruitment years resulted from high reproduction in 1980/1981 and 2000/2001. Elevated recruitment from 2000/2001 larvae also was observed downstream at South Georgia (Siegel et al., 2003).

In contrast, conditions during 2001/2002 were not as favorable for a successful reproduction, as evidenced by the lower number of larvae recorded during fall 2002 (this study; Daly, 2004) and the subsequent low numbers of krill observed during January 2003 along the Peninsula (Ducklow et al., 2006).

Results reported herein suggest that seasonal persistence of elevated food concentrations from phytoplankton blooms during spring and summer likely were a strong influence on krill reproduction and recruitment, particularly in 2001. Recruitment also may vary along the Peninsula depending on the location of phytoplankton blooms. For example during 1999/2000, higher chlorophyll concentrations occurred in the Elephant Island - Scotia Sea areas relative to any other year analyzed (Fig. 3.4b: regions 3, 6, and 12). Published krill recruitment index values indicate that elevated recruitment occurred in 2001 from the 1999/2000 larvae ($R_1 = 0.573$) in this area (Siegel et al., 2002). On the other hand, chlorophyll concentrations in the southern sectors of the western Antarctic
Peninsula were average or below-average in coastal waters including Marguerite Bay early in the season (regions 7, 8, 9, 10, 11, 13), although by February an increase could be observed in most regions along the continental shelf. Consistent with the lower availability of food during the critical early reproductive period, recruitment indices in the southern part of the study area for the 1999/2000 larvae were low \([R_1 = 0.076\] in summer; (Siegel et al., 2003) and \(R_1 = 0\) in fall (this study)]).

Chlorophyll concentrations in the Bellingshausen may have influenced downstream densities of larval krill in the vicinity of Marguerite Bay, especially at the offshelf station in the ACC. Recent modeling studies suggest that krill spawned in the Bellingshausen Sea are transported downstream to the Western Antarctic Peninsula area and into the Scotia Sea (e.g., Thorpe et al., 2007). The wide range of larval stages observed in offshelf waters during fall 2001 indicated that krill reproduction started relatively early in the season and continued for an extended period. The dominant larval modes included CIII, FI, and FII, in addition to considerable numbers of late stage furcilia. Based on experimentally determined growth rates (Ikeda, 1984), these larvae represent a range of spawning episodes between mid-December (FVI: ~ 127 days old) and early to late-March (CII: ~ 44 days old). Assuming that most reproducing adult females released eggs in the vicinity of the shelf break (Siegel, 1988), the approximate location of the spawning population upstream in the ACC may be estimated from the age of larvae and the transport rate of the current. For example, the dominant FI mode in 2001 is estimated to be about 63 days old and, therefore, likely originated from a late February - early March reproductive event. Surface velocities in the ACC reach 0.25 - 0.4 m sec\(^{-1}\), but decrease monotonically with depth (Klinck and Nowlin, 2001). Mesoscale
meanders and eddies also may act to reduce the transport rate. Assuming an average eastward current velocity of the ACC of \(\sim 0.1 - 0.2 \text{ m sec}^{-1}\), the spawning location possibly occurred in offshelf waters of the Bellingshausen Sea between 83 - 94 °W (Fig. 3.8; white box). Although this spawning location is west of our study area, larger scale ocean color imagery (Fig. 3.8a) shows large blooms in this region in 2001, which would provide adequate food for an early and extended reproduction by adults and provide food for larvae transported to the east. In contrast during 2002 chlorophyll concentrations during summer were relatively low (Fig. 3.8b) and, thus, there was less food available for reproduction or larval growth.

The results obtained during the present study indicate that spring and summer phytoplankton blooms appear to be a significant factor influencing krill recruitment in the vicinity of the Antarctic Peninsula. Clearly, further studies of the factors controlling phytoplankton blooms in waters adjacent to the southern Antarctic Peninsula and in the Bellingshausen Sea are warranted, especially since this area may play a major role in krill reproduction and influence other components of the Antarctic food web. In addition, further physiological-based krill studies are needed to better understand the relative impact of summer phytoplankton blooms versus winter sea ice cover in governing recruitment, particularly in light of the regional decline in sea ice.
Figure 3.8. Monthly mean SeaWiFS chlorophyll concentrations in the Bellingshausen Sea during February (a) 2001 and (b) 2002. The white box represents the area of potential krill spawning. The red star indicates the location of the US SO GLOBEC sampling Station 1 where krill were collected. Images correspond to the level-3 standard mapped images (resolution of 9 km/pixel) and were obtained from the global ocean color imagery (http://oceancolor.gsfc.nasa.gov/cgi/level3.pl).
CHAPTER 4

PHYSICAL AND BIOLOGICAL CONTROLS ON INTERANNUAL
VARIABILITY OF ZOOPLANKTON IN MARGUERITE BAY, WESTERN
ANTARCTIC PENINSULA, AUSTRAL FALL 2001 AND 2002

INTRODUCTION

Zooplankton are a key component of marine ecosystems, acting as the link between primary producers and higher trophic levels. In the Southern Ocean, the Antarctic krill, *Euphausia superba*, often dominates zooplankton biomass and is considered a keystone species. It is a relatively large euphausiid, up to 65 mm in length, and an important phytoplankton grazer, particularly during spring and summer, and prey for many upper trophic level predators, including fish, penguins, seals, and whales. *E. superba* has a circumpolar distribution and in deep water can often be observed in dense aggregations over the continental shelf. Due to its vital role in the Antarctic ecosystem, the zooplankton literature is dominated by studies of this species. Extensive research has focused on examining the distribution and abundance of *E. superba* (e.g., Marr, 1962; Siegel, 1992), its reproduction and growth (Ross and Quetin, 1983; Brinton et al., 1986; Daly, 1990; Quetin et al., 2003), behavior (e.g., Hamner et al., 1983; Marschall, 1988) and role in the Antarctic foodweb (Hempel, 1985; Laws, 1985). In addition to this species, other zooplankton, including other euphausiid species and copepods, are
important components of the Antarctic ecosystem. Nonetheless, the information available on their role in the ecosystem, population dynamics, and ecology is limited compared to that of *E. superba*.

*Thysanoessa macrura*, an omnivorous euphausiid up to ~29 mm in length (Nordhausen, 1992), is broadly distributed in coastal and oceanic waters of the Southern Ocean and at all temperature and salinity ranges (Hempel and Marschoff, 1980). Its distribution overlaps that of *E. superba* and both species often co-occur. The larvae of *T. macrura* are herbivorous and graze on phytoplankton during spring and summer, whereas the adults prey mainly on copepods (Hopkins, 1985). *E. crystallorophias* is another relatively small euphausiid (up to ~36 mm) found exclusively in neritic waters, where it either co-exists or replaces *E. superba* (Daly and Zimmerman, 2004). *E. crystallorophias* is mainly herbivorous throughout its life cycle. Both *E. crystallorophias* and *T. macrura* can be a significant food item for higher trophic level predators, including penguins, fish and whales and, thus, are also important members of the Antarctic foodweb (Croxall, 1984; Nemoto and Nasu, 1958; Ainley and DeMaster, 1990).

Copepods dominate zooplankton abundance in many regions of the Southern Ocean and their biomass may exceed that of krill at times (Schnack-Schiel and Mujica, 1994). The dominant species include the herbivore *Calanoides acutus* and several omnivorous species, including *Metridia gerlachei*, *Calanus propinquus* and *Ctenocalanus* spp. Several studies have analyzed the patterns of distribution and abundance of the dominant copepods in different areas of the Southern Ocean and have emphasized their critical role in the zooplankton community, both as prey for carnivorous zooplankton and fish, and also as predators of phytoplankton and microzooplankton (e.g.
Hopkins, 1985; Conover and Huntley, 1991; Schnack-Schiel et al., 1991; Lopez and Huntley, 1995; Atkinson, 1998; Voronina, 1998; Schnack-Schiel, 2001). In addition, other zooplankton, such as *E. superba*, have been observed to rely on copepods during fall and winter when phytoplankton is scarce in the water column (Hopkins, 1985; Daly, *pers. comm.*).

The western Antarctic Peninsula (WAP) is one of the most productive areas of the Southern Ocean, supporting high concentrations of phytoplankton, zooplankton and upper trophic level predators (Deibel and Daly, 2007). This area also is of interest due to its rapid warming relative to any other area in the world’s ocean (Vaughan and Doake, 1996) and the dramatic decline in sea ice cover observed in recent decades (Parkinson, 2002), which in turn has important implications for Antarctic organisms. Although elevated chlorophyll concentrations have been observed along the continental shelf of the northern WAP during austral spring and summer (Moline et al., 1997; Smith et al., 1998a; Garibotti et al., 2003), recent results have shown that phytoplankton blooms are particularly large and persistent in the southeastern sectors, including Marguerite Bay and the western Bellingshausen Sea (Chapter 3; Marrari et al., 2008). The factors that contribute to the relatively high productivity of this region include its northern location relative to other Antarctic shelf areas, a relatively wide continental shelf, and the intrusion onto the shelf of nutrient-rich Upper Circumpolar Deep Water (UCDW) (Deibel and Daly, 2007). Numerous studies also have examined zooplankton in the WAP region, which have improved our understanding of the patterns of abundance, distribution and reproduction of the dominant taxa (e.g. Hopkins, 1985; Siegel, 1988; 1992; Mujica, 1989; Schnack-Schiel and Mujica, 1994; Ross et al., 1996; Siegel and Harm, 1996; Loeb et al., 1996).
However, these studies almost exclusively took place in the northernmost sectors of the Peninsula, from Adelaide Island toward the northeast, and only a few extended further south to the outer shelf off Marguerite Bay (Lascara et al., 1999, Meyer et al., 2003). Despite the critical role of the southern sector in the ecosystem of the WAP, studies of zooplankton from Marguerite Bay and the Bellingshausen Sea have been scarce (Atkinson, 1995; Siegel and Harm, 1996; Meyer et al., 2003).

The Southern Ocean Global Ocean Ecosystem Dynamics Program (SO GLOBEC) focused its study in the vicinity of Marguerite Bay, as this region was believed to be an important overwintering habitat for *E. superba* based on observations of predators during winter (Hofmann et al., 2004). The main objective of SO GLOBEC was to investigate the physical and biological factors that influence the growth, recruitment and overwintering survival of *E. superba* (Hoffman et al., 2004). The program supported two ships operating simultaneously during two fall and winter cruises in the vicinity of Marguerite Bay in 2001 and 2002. In addition to krill data, an extensive dataset involving other important zooplankton groups was generated from net samples.

Marguerite Bay should be a favorable environment for zooplankton owing to persistent elevated concentrations of phytoplankton, the availability of protected areas such as fjords and bays that serve as refuge from advection out of the area, and the intrusion of UCDW onto the shelf through Marguerite Trough, a deep canyon (> 500 m) that intercepts the continental shelf break off Marguerite Bay. In addition, the flow of the Antarctic Peninsula Coastal Current (APPC) (Beardsley et al., 2004; Klinck et al., 2004; Moffat et al., 2008) in this area involves a gyre-like feature that contributes to making Marguerite Bay a favorable retention area for both phyto- and zooplankton, and thus a
potentially favorable feeding ground for predators.

Understanding the dynamics of zooplankton populations and how they respond to environmental change is critical to assessing the impact that these changes will have on the Antarctic ecosystem as a whole, and in particular on upper trophic level predators that rely on zooplankton as a food source. Herein, I investigate the patterns of abundance and distribution of dominant zooplankton in Marguerite Bay during austral fall 2001 and 2002 in relation to the interannual variability of environmental conditions, and examine possible ecological relationships within and between groups.

METHODS

The study area consisted of coastal waters in the vicinity of Marguerite Bay along the western Antarctic Peninsula region between 66 - 70 °S and 67 - 73 °W. Zooplankton and environmental data were collected between 23 April and 6 June 2001 and 7 April and 20 May 2002 during process cruises onboard the R.V. Lawrence M. Gould, as part of the SO GLOBEC Program. The process cruises occupied six coastal stations and samples from a total of 12 zooplankton net hauls were obtained during each cruise (Fig. 4.1). In addition, Video Plankton Recorder data of zooplankton were obtained aboard the R.V. Nathaniel B. Palmer during survey cruises along 13 transects spaced 40 km apart on the mid to outer shelf off Marguerite Bay (Ashjian et al., 2008). Results reported here are derived from net samples obtained during survey cruises.

Zooplankton samples were collected at eight discrete depth intervals using a 1m² Multiple Opening-Closing Net System and Environmental Sensing System (MOCNESS), having 333 μm mesh. Maximum sampling depths ranged between 200 and 800 m,
depending on bathymetry. Samples were preserved in 10% formalin and stored for analysis in the laboratory. Samples were initially split to include approximately 100 individuals of the dominant euphausiid species. For copepod counts and identification, samples were split further to include approximately 100 individuals of the dominant copepod. All zooplankton taxa present in the subsamples were identified and counted. Euphausiids were identified to species and developmental stage (larval stages, juveniles, adult females and adult males) after Makarov (1981) and Mauchline (1981), and measured for total length to the nearest half mm (from the base of the eye to the tip of the telson, excluding setae). For plotting purposes, length data were grouped in 1 mm length bins. Only data for juveniles > 20 mm total length and adults are included in this study. Copepods were identified for species and enumerated. For Euchaetidae, the designation of Park (1994) was followed, who ascribed the Antarctic species to the genus *Paraeuchaeta*. All other zooplankton groups were identified, counted, measured, and classified into > 15 mm or < 15 mm total length; however, analyses in this study combined the abundances of these size categories.
Figure 4.1. (a) Location of the study area (red rectangle) and MOCNESS net hauls (circles) during fall of (b) 2001 and (c) 2002. General station locations and geographic references are also indicated. MB is Marguerite Bay.

Water column integrated abundance \((A, \text{ind m}^{-2})\) was calculated for each taxa in all net hauls using:

\[
A = \sum_{i=1}^{8} n_i z_i
\]

where \(n\) is abundance (ind m\(^{-3}\)) in net \(i\), and \(z\) is the depth interval (m) of the stratum sampled by net \(i\), calculated as the difference between the depth at which the net was
closed and the depth at which it was opened (e.g., if stratum sampled was 100 - 150 m, z is 50 m). A mean water column integrated abundance was estimated for each taxa during each year (n = 12). In addition, a mean integrated abundance was calculated for all taxa at each station, when multiple net hauls were done at a station.

The depth of maximum abundance (Z) was obtained for each taxa in each net haul by calculating the center-depth of the stratum with the highest abundance at a given location. A mean depth of maximum abundance was calculated for each taxa during each year.

Abundances and depth distributions for all taxa were tested for normality (Shapiro-Wilk W Test) (Shapiro et al., 1968). Abundances (A) and depths of maximum abundance (Z) for all dominant copepod species followed a normal distribution; therefore, statistical analyses for copepod parameters included arithmetic means, Student’s t-Test, and one-way ANOVA at α = 0.05 (Zar, 1984). Abundances and vertical distributions of all other groups, including euphausiids, were not normally distributed; thus, statistical analyses reported here are nonparametric and included geometric means, Mann-Whitney U Test, and Kruskall-Wallis ANOVA at α = 0.05 (Zar, 1984).

Vertical profiles of conductivity, temperature, and density were obtained with a CTD mounted on the MOCNESS during all net hauls. In addition, water column chlorophyll samples were collected at several CTD stations in proximity to the MOCNESS locations. A total of 15 and 6 casts are available for fall 2001 and 2002, respectively. At each CTD station, chlorophyll samples for 4 - 9 depths were collected using 10-L Niskin bottles mounted on a rosette. Water was filtered onto GF/F filters and pigments were extracted in 90% acetone at -20 °C in the dark for at least 24 hours.
Chlorophyll fluorescence was measured on a Turner Design Digital 10-AU-05 fluorometer calibrated prior to each cruise. MOCNESS tows were matched to the geographically closest CTD cast for comparisons between zooplankton and environmental data (Fig. 4.1). Spearman rank order correlations were calculated between the vertically integrated abundance of all zooplankton taxa and several environmental variables: vertically integrated pigment concentrations (chlorophyll + phaeopigment; mg m$^{-2}$), bottom depth (m), and salinity at 10 m.

In addition to the concurrent environmental variables measured during fall, monthly mean chlorophyll concentrations between 45 - 75 °S and 50 - 80 °W were obtained from SeaWiFS for the preceding summer season. More detail on the processing of SeaWiFS data are presented in Chapters 2 and 3. Analyses in this chapter include five of the subregions defined in Chapter 3 (Fig. 3.4a), which represent oceanic and coastal waters of the Bellingshausen Sea and Marguerite Bay. The geometric mean chlorophyll for the five subregions was calculated for each biweekly period from October - April during spring-summer 2000/2001 and 2001/2002. Data are presented in relation to the seven-year climatology calculated as the geometric mean for 1997 - 2004 at each subregion.

RESULTS

*Abundance and Percent Contribution*

The zooplankton of Marguerite Bay comprised 13 major taxonomic groups,
including 12 species of copepods, three species of euphausiids, and 10 other taxa (Tables 4.1 and 4.2). The dominant copepods included Calanoides acutus, Metridia gerlachei, Ctenocalanus spp., and Paraeuchaeta spp. Oithona spp., Oncaea spp., Calanus propinquus, Rhincalanus gigas, Gaidius tenuispinus and Scolecithricella minor were present in generally lower numbers. Aetidopsis antarctica and Metridia curticauda were present only occasionally in low abundances and, therefore, were excluded from the remaining analyses. Overall, copepods were significantly more abundant in fall 2001 (24,362 ± 11,948 ind m⁻²) than 2002 (9,496 ± 6,450 ind m⁻²) (Student’s t-Test; p < 0.001) (Fig. 4.2a). During 2001, abundances were highest in Laubeuf Fjord (Sta. 5), at the northern end of Alexander Island (Sta. 4a), George VI Sound (Sta. 4b), Lazarev Bay (Sta. 3), and Neny Fjord (Sta. 6). In 2002, copepods were most abundant in the vicinity of Alexander Island and southern Laubeuf Fjord. Densities were lowest in Crystal Sound (Sta. 7), Bourgeois Fjord (Sta. 5), and at the southern end of Adelaide Island (Avian Island). Abundances also were low in northern Laubeuf Fjord during 2002. In addition, copepod species percent composition showed interannual differences (Fig. 4.2b). In 2001, C. acutus and M. gerlachei dominated the community (mean = 52 and 38% of the total copepod abundance, respectively), followed by Paraeuchaeta spp. (5%). In contrast, during 2002 the number of relatively abundant species (> 5%) was higher and included M. gerlachei (32%) Ctenocalanus spp. (27%), C. acutus (18%), Oithona spp. (10%), and Paraeuchaeta spp. (8%). C. propinquus and R. gigas, two relatively large species frequently found in high abundances in the Southern Ocean, only accounted for < 3% of total copepod abundance during both years. Oncaea spp., a frequently dominant small copepod (< 1 mm) was generally present in low abundances throughout the study.
area, and only accounted for 1.4% and 2.6% of total copepods during 2001 and 2002, respectively.

Table 4.1. Copepod abundance (ind m\(^{-2}\)) in the vicinity of Marguerite Bay during austral fall 2001 and 2002. Mean = arithmetic mean, SD = standard deviation, n = number of net hauls in which a species was present.

<table>
<thead>
<tr>
<th>Taxonomic group</th>
<th>Fall 2001</th>
<th></th>
<th></th>
<th>Fall 2002</th>
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</tr>
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<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>n</td>
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</tr>
<tr>
<td>Copepods</td>
<td></td>
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<td>Aetidopsis antarctica</td>
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<td>37.9</td>
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<td>6.29</td>
<td>21.8</td>
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<td>12</td>
<td>1909</td>
<td>1891</td>
<td>12</td>
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<td>145</td>
<td>121</td>
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<td>173</td>
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<td>11</td>
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<td>12</td>
<td>2497</td>
<td>1856</td>
<td>11</td>
</tr>
<tr>
<td>Gaidius tenuispinus</td>
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<td>75.9</td>
<td>9</td>
<td>32.9</td>
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<td>7</td>
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<tr>
<td>Metridia curticauda</td>
<td>0</td>
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<td>0</td>
<td>7.87</td>
<td>27.3</td>
<td>1</td>
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<td>5487</td>
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<td>3153</td>
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<td>994</td>
<td>12</td>
</tr>
<tr>
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<td>12</td>
<td>239</td>
<td>264</td>
<td>10</td>
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<td>Paraeuchaeta spp.</td>
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<tr>
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<td>30.0</td>
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<tr>
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<td>13.2</td>
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</tbody>
</table>
Table 4.2. Zooplankton abundance (ind m$^{-2}$) in the vicinity of Marguerite Bay during austral fall 2001 and 2002. Geo mean = geometric mean, Range = minimum - maximum, n = number of net hauls in which a taxon was present

<table>
<thead>
<tr>
<th>Taxonomic group</th>
<th>Geo mean</th>
<th>Range</th>
<th>n</th>
<th>Geo mean</th>
<th>Range</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ostracods</td>
<td>775</td>
<td>32.4 - 4728</td>
<td>12</td>
<td>788</td>
<td>83.6 - 2517</td>
<td>12</td>
</tr>
<tr>
<td>Euphausiids</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Euphausia crystallorophias</td>
<td>2.78</td>
<td>0.24 - 13.2</td>
<td>10</td>
<td>18.5</td>
<td>1.93 - 137</td>
<td>12</td>
</tr>
<tr>
<td>Euphausia superba</td>
<td>6.22</td>
<td>0.24 - 72.6</td>
<td>11</td>
<td>14.8</td>
<td>1.63 - 117</td>
<td>11</td>
</tr>
<tr>
<td>Thysanoessa macrura</td>
<td>45.9</td>
<td>7.19 - 266</td>
<td>12</td>
<td>16.2</td>
<td>4.27 - 51.9</td>
<td>12</td>
</tr>
<tr>
<td>Amphipods</td>
<td>15.1</td>
<td>5.63 - 91.5</td>
<td>12</td>
<td>12.7</td>
<td>6.77 - 86.6</td>
<td>12</td>
</tr>
<tr>
<td>Mysids</td>
<td>1.73</td>
<td>0.23 - 34.9</td>
<td>11</td>
<td>4.26</td>
<td>0.75 - 29.8</td>
<td>10</td>
</tr>
<tr>
<td>Medusae</td>
<td>13.3</td>
<td>0.27 - 520</td>
<td>9</td>
<td>0.39</td>
<td>0.12 - 1.47</td>
<td>6</td>
</tr>
<tr>
<td>Siphonophores</td>
<td>present</td>
<td>-</td>
<td>12</td>
<td>present</td>
<td>-</td>
<td>11</td>
</tr>
<tr>
<td>Polychaetes</td>
<td>136</td>
<td>0.65 - 1110</td>
<td>10</td>
<td>11.9</td>
<td>0.48 - 279</td>
<td>11</td>
</tr>
<tr>
<td>Pteropods</td>
<td>254</td>
<td>2.33 - 2119</td>
<td>12</td>
<td>273</td>
<td>7.63 - 1556</td>
<td>11</td>
</tr>
<tr>
<td>Chaetognaths</td>
<td>57.0</td>
<td>3.05 - 687</td>
<td>12</td>
<td>31.3</td>
<td>4.23 - 161</td>
<td>12</td>
</tr>
<tr>
<td>Appendicularians</td>
<td>4.46</td>
<td>0.78 - 32.3</td>
<td>3</td>
<td>1447</td>
<td>147 - 5693</td>
<td>4</td>
</tr>
<tr>
<td>Salps</td>
<td>3.22</td>
<td>0.61 - 43.9</td>
<td>3</td>
<td>0.22</td>
<td>0.15 - 0.27</td>
<td>3</td>
</tr>
</tbody>
</table>
Figure 4.2a. Water column integrated abundance (ind m$^{-2}$) of copepods from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom).
Figure 4.2b. Percent composition of copepods at coastal stations in Marguerite Bay during austral fall 2001 (top) and 2002 (bottom). Color legend as in Figure 4.2a.
The overall geometric mean abundance of total euphausiids was very similar during both years: 66.3 and 66.8 ind m$^{-2}$ in 2001 and 2002, respectively (Mann Whitney U; $p = 1.00$) (Fig. 4.3a). Although abundances were variable within sampling locations and over the study area, distribution patterns were consistent between years. Northern areas, such as Laubeuf Fjord and Crystal Sound, consistently had the highest integrated total euphausiid abundances. The southern sectors, including Neny Fjord, inner Marguerite Bay and near Alexander Island, had relatively lower euphausiid abundances during both years. George VI Sound and Lazarev Bay were only sampled during 2001 and had the lowest abundances of all areas surveyed.

Despite the similarity in total euphausiid abundance and distribution between years, there were significant interannual differences in species percent composition (Fig. 4.3b). *T. macrura* was the most abundant species during 2001 with a geometric mean abundance of 45.9 ind m$^{-2}$, whereas *E. crystallorophias* had overall low abundances (2.78 ind m$^{-2}$), and *E. superba* had intermediate values (6.22 ind m$^{-2}$). During fall 2002, abundances were similar among species, with *E. crystallorophias* having the highest mean value (18.5 ind m$^{-2}$), followed by *T. macrura* (16.2 ind m$^{-2}$), and finally *E. superba* with the lowest abundance (14.8 ind m$^{-2}$). Interannual differences in abundances of *T. macrura* (Mann Whitney U; $p = 0.020$) and *E. crystallorophias* (Mann Whitney U; $p = 0.004$) were significant, whereas values for *E. superba* were not significantly different between years (Mann Whitney U; $p = 0.200$).
Figure 4.3a. Water column integrated abundances of euphausiids (ind m$^{-2}$) from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom).
Figure 4.3b. Percent composition of euphausiids from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom). Color legend as in Figure 4.3a.
Even though abundances of *E. superba* were comparable between years, there were key interannual differences in developmental stage composition (Fig. 4.4). During 2001, non-larval *E. superba* were almost exclusively adult males and females and juveniles were essentially absent. Only a few year-two juveniles, 34 mm total length (TL), were observed at one station in Laubeuf Fjord (0.08 ind m$^{-3}$). Overall, juveniles accounted for only ~2% of the postlarval *E. superba* densities in 2001. In contrast, year-one juveniles were abundant and constituted ~41% of the populations in 2002, suggesting a successful recruitment from 2001 larvae. Postlarval *T. macrura* were mostly juveniles 8 - 10 mm TL during 2001 and 2002 (Fig. 4.5). Juveniles comprised 91% of individuals in fall 2001, and ~69% in 2002, when the remaining 31% were adults ranging primarily between 16 - 20 mm TL. During 2001, *E. crystallorophias* juveniles (10 - 18 mm TL) comprised 11% of the population, while the remaining fraction (89%) included adults 20 - 36 mm TL (mode: 29 mm TL) (Fig. 4.6). In 2002, juveniles were a larger fraction of the population (28%), while the adults had a bimodal distribution, with a larger mode at 20 mm TL, and lower proportions of larger individuals between 29 - 33 mm TL. Even though the percent composition of postlarval *E. crystallorophias* was comparable between years, larval abundances showed marked interannual differences, with higher values in 2001 throughout the study area. Maximum abundances in 2001 were 829 ind m$^{-2}$ in Neny Fjord, whereas the highest values recorded in 2002 were ~18 ind m$^{-2}$ in northern Laubeuf Fjord (data not shown).

In addition to copepods and euphausiids, other dominant zooplankton groups included ostracods, pteropods, polychaetes, chaetognaths, appendicularians, amphipods, and mysids (Fig. 4.7a and b). Ostracods and pteropods were numerically dominant
during both years and showed no significant interannual differences (Mann Whitney U
test, p = 1.00 for ostracods, p = 0.805 for pteropods). Appendicularians were more
abundant during 2002 (geometric mean = 1447 ind m⁻²) although their distribution was
very patchy and they were present in only four net hauls. During 2001, they were present
at three locations, but in very low abundances. Polychaetes and chaetognaths were
relatively abundant during 2001, but had lower densities in 2002. Abundances for
amphipods and mysids were not significantly different between years (Mann Whitney U
test, p = 0.32 for amphipods, p = 0.204 for mysids). Gelatinous zooplankton were
generally rare. Siphonophores could not be numerically quantified due to the presence of
only fragments of colonies in the samples; however, fragments were classified as “few”
or “numerous” and were frequent throughout the water column at most locations during
both years, particularly in the upper 300 m.

Laubeuf Fjord had the highest abundances of these groups of zooplankton,
followed by the areas in the vicinity of Alexander Island and Neny Fjord/inner
Marguerite Bay (Sta. 6) during both years (Fig. 4.7a). In 2001, abundances were lowest
in Bourgeois Fjord, one station in southern George VI Sound, and Lazarev Bay, while
during the following fall, densities were lowest in Crystal Sound and south of Adelaide
Island. In terms of percent composition, during 2001 ostracods, pteropods and
polychaetes dominated at all stations except in Lazarev Bay. At this location, these
groups were scarce or absent, and the community was composed primarily of medusae,
chaetognaths, and salps (Fig. 4.7b). Although ostracods dominated the zooplankton
numerically at most locations in 2001, their percent contribution varied among stations.
Their contribution was maximum in Crystal Sound, Laubeuf Fjord, and George VI
Sound, and minimum in the vicinity of Alexander Island and Lazarev Bay. In 2002, ostracods again dominated numerically at most locations, but were only a minor fraction of zooplankton in some net hauls in Laubeuf Fjord, Adelaide Island, and Marguerite Trough, where appendicularians were most important.
Figure 4.4. Length-frequency of *E. superba* juveniles and adults during fall 2001 (top) and 2002 (bottom). Data represent all coastal net hauls for each year. Only juveniles $\geq 20$ mm are included.
Figure 4.5. Length-frequency of *T. macrura* juveniles and adults during fall 2001 (top) and 2002 (bottom). Data represent all coastal net hauls for each year.
Figure 4.6. Length-frequency of *E. crystallorophias* juveniles and adults during fall 2001 (top) and 2002 (bottom). Data represent all coastal net hauls for each year.
Figure 4.7a. Water column integrated abundances of zooplankton other than copepods and euphausiids (ind m$^{-2}$) from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom).
Figure 4.7b. Percent composition of zooplankton other than copepods and euphausiids from net hauls in the vicinity of Marguerite Bay during austral fall 2001 (top) and 2002 (bottom). Color legend as in Figure 4.5a.
**Vertical Distribution**

Overall, the mean depth of maximum abundance for all copepods was significantly deeper in the water column during May - June 2001, than in April - May 2002 (2001: 252 m; 2002: 75 m; Student’s t-Test; p < 0.001) (Fig. 4.8). Looking at individual species, only depths of *Paraeuchaeta* spp., *Oithona* spp., and *R. gigas* were significantly different between years (ANOVA, p < 0.05), although other species such as *M. gerlachei*, *Oncaea* spp., and *S. minor* also had somewhat shallower distributions in 2002. Copepod species occurred within three depth ranges: shallow (0 - 100 m), intermediate (100 - 150 m), and deep (> 150 m) water groups (Table 4.3). In 2001, shallow species included *Ctenocalanus* spp. and *C. propinquus*, while *Oithona* spp., *S. minor*, *Paraeuchaeta* spp., *M. gerlachei*, *C. acutus*, *G. tenuispinus* and *Oncaea* spp. were identified as deep species. Only *R. gigas* was intermediate during the first year. In contrast, *Oithona* spp., *R. gigas*, and *Paraeuchaeta* spp. were grouped as shallow species during 2002, while *M. gerlachei* and *S. minor* were intermediate. *Oncaea* spp., *C. acutus*, and *G. tenuispinus* remained deep species during fall 2002.

The vertical distributions of euphausiids were similar within and between years (Table 4.4) and geometric mean depths of maximum abundance for all species were in the upper 100 m (65 - 84 m). Mysids were distributed the deepest during 2001 (249 m) and 2002 (224 m), while amphipods were located at intermediate depths in 2001 (125 m) and represented the shallowest group during 2002 (55 m). The depths of maximum abundance for *E. superba*, *E. crystallorophias*, and *T. macrura* were not significantly different within or between years (Kruskall-Wallis ANOVA; p = 0.882 in 2001, and p = 0.873 in 2002). However, even though their general depth ranges overlapped throughout
Table 4.3. Mean depth of maximum abundance (Z, m) of copepods in 2001 and 2002. Species in bold indicate those that were classified in the same depth category during both years. The range represents the shallowest and deepest depth (m) at which each species was present. Depth categories were based on the general hydrography of the area. At most net hauls, the upper mixed layer ranged between 0 - ~100 m, with a thermocline/pycnocline generally between ~100 - 150 m, and warmer saltier water below ~150 m.

<table>
<thead>
<tr>
<th></th>
<th>Fall 2001</th>
<th>Fall 2002</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Shallow</strong></td>
<td><strong>Z</strong></td>
<td><strong>Range</strong></td>
</tr>
<tr>
<td><strong>Ctenocalanus spp.</strong></td>
<td>51</td>
<td>0 - 500</td>
</tr>
<tr>
<td><strong>C. propinquus</strong></td>
<td>80</td>
<td>0 - 525</td>
</tr>
<tr>
<td><strong>Intermediate</strong></td>
<td><strong>Z</strong></td>
<td><strong>Range</strong></td>
</tr>
<tr>
<td><strong>R. gigas</strong></td>
<td>147</td>
<td>0 - 800</td>
</tr>
<tr>
<td><strong>M. gerlachei</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>S. minor</strong></td>
<td>184</td>
<td>0 - 350</td>
</tr>
<tr>
<td><strong>G. tenuispinus</strong></td>
<td>230</td>
<td>0 - 500</td>
</tr>
<tr>
<td><strong>Deep</strong></td>
<td><strong>Z</strong></td>
<td><strong>Range</strong></td>
</tr>
<tr>
<td><strong>Oithona spp.</strong></td>
<td>162</td>
<td>0 - 525</td>
</tr>
<tr>
<td><strong>S. minor</strong></td>
<td>184</td>
<td>0 - 350</td>
</tr>
<tr>
<td><strong>Paraeuchaeta spp.</strong></td>
<td>201</td>
<td>0 - 800</td>
</tr>
<tr>
<td><strong>C. acutus</strong></td>
<td>213</td>
<td>0 - 800</td>
</tr>
<tr>
<td><strong>G. tenuispinus</strong></td>
<td>249</td>
<td>0 - 800</td>
</tr>
<tr>
<td><strong>Oncaea spp.</strong></td>
<td>348</td>
<td>0 - 800</td>
</tr>
</tbody>
</table>
the study area, the depths of maximum abundance of species usually did not overlap at any given location (Fig. 4.8). For example in Crystal Sound during 2001, the maximum abundances of *E. superba* occurred at 50 m, while *T. macrura* primarily occurred at 100 m (Fig. 4.9a). In 2002, the depth of maximum abundance for *E. superba* was 150 m, *E. crystallorophias* was primarily found at 50 m, and *T. macrura* was at 75 m. In Laubeuf Fjord, *E. superba* and *T. macrura* had peaks at 100 m in 2001, while *E. crystallorophias* was observed at 50 m, and a second mode for *T. macrura* occurred at 400 - 500 m (Fig. 4.9b). In 2002, *E. superba* had peaks of maximum abundance at 75 m and 150 m, while
*E. crystallorophias* occurred primarily at 100 m. In the vicinity of Alexander Island during fall 2001, *E. superba* could be found mostly at 50 m, while *T. macrura* was mainly at 100 m (Fig. 4.9c). Finally, south of Adelaide Island, elevated abundances of *E. superba* were located at shallow depths in 2002 (0 - 100 m, maximum at 50 m), while *T. macrura* showed a smaller peak at 150 m, and *E. crystallorophias* was not present in significant numbers.

The geometric mean depths of maximum abundances of all euphausiid species ranged between 65 - 73 m in 2001, while the majority of copepods were located at 252 m during the same year. Looking at individual net hauls for 2001, the maximum abundances of both groups did not overlap at almost any location, with the bulk of the copepod community located consistently deeper than the mode of the euphausiids (Appendix 1). On the other hand, during 2002, euphausiids were observed at similar depths as the previous fall, with maximum abundances between 70 - 84 m (Table 4.4), while copepods were significantly shallower than in 2001, mostly at 75 m (Appendix 2).

### Table 4.4

Mean depth of maximum abundance (m) of euphausiids, amphipods, and mysids during fall 2001 and 2002. Data include all coastal stations for each cruise. Values represent the geometric mean depth of maximum abundance of each group for all net hauls during each year (n = 12). The range (m) represents the range of depths in the water column that were occupied by each taxon.

<table>
<thead>
<tr>
<th></th>
<th>Fall 2001</th>
<th>Fall 2002</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Depth (m)</td>
<td>Range</td>
</tr>
<tr>
<td><em>E. crystallorophias</em></td>
<td>65</td>
<td>0 - 500</td>
</tr>
<tr>
<td><em>E. superba</em></td>
<td>73</td>
<td>0 - 500</td>
</tr>
<tr>
<td><em>T. macrura</em></td>
<td>68</td>
<td>0 - 800</td>
</tr>
<tr>
<td>Amphipods</td>
<td>125</td>
<td>0 - 800</td>
</tr>
<tr>
<td>Mysids</td>
<td>249</td>
<td>0 - 500</td>
</tr>
</tbody>
</table>
Figure 4.9a. Vertical distribution of euphausiids, amphipods, and mysids (ind m$^{-3}$) in Crystal Sound during austral fall 2001 (top) and 2002 (bottom).
Figure 4.9b. Vertical distribution of euphausiids, amphipods, and mysids (ind m$^{-3}$) in Laubeuf Fjord during austral fall 2001 (top) and 2002 (bottom).
Figure 4.9c. Vertical distribution of euphausiids, amphipods, and mysids (ind m\(^{-3}\)) in the vicinity of Alexander Island during austral fall 2001 (top) and south of Adelaide Island in fall 2002 (bottom).
Horizontal Distribution

The horizontal distribution of zooplankton varied within the study area. In general, in areas where euphausiids, amphipods, and mysids (i.e. macrozooplankton) were abundant, copepod densities were low, and vice versa (Fig. 4.10a).

Macrozooplankton were most abundant in northern sectors, whereas copepod abundances were highest in southern sectors, such as inner Marguerite Bay, the vicinity of Alexander Island, George VI Sound and Lazarev Bay (Fig. 4.10b). Exceptions to this pattern were observed in fall 2001 in northern Laubeuf Fjord, where copepods and macrozooplankton were highly abundant, in Bourgeois Fjord, where all zooplankton were scarce, and south of Adelaide Island in 2002, where copepod and macrozooplankton abundances were low.

Figure. 4.10a. Linear correlation between integrated abundances (ind m\(^{-2}\)) of total macrozooplankton (euphausids, amphipods, and mysids) and copepods at different stations during fall 2001 and 2002 (n = 12; r = -0.39; p = 0.208). When multiple net hauls were sampled, values represent the mean. Plot includes data for stations in Crystal Sound (Sta. 7), Laubeuf and Bourgeois fjords (Sta. 5), Neny Fjord/inner Marguerite Bay (Sta. 6), Marguerite Trough (Sta. 2), S. Adelaide Island (Avian Is.), Alexander Island (Sta. 4a), George VI Sound (Sta. 4b), and Lazarev Bay (Sta. 3). Data for Laubeuf Fjord in 2001 were excluded (outlier).
Figure 4.10b. Mean water column integrated abundance (ind m$^{-2}$) of macrozooplankton (euphausiids, amphipods, and mysids) (grey bars, left axis) and copepods (black circles, right axis) at different stations within Marguerite Bay during fall 2001 (top) and 2002 (bottom).
Fall Environmental Parameters

The environmental parameters investigated here were chosen based on their potential influence on zooplankton distributions: vertically integrated pigment concentrations (chlorophyll + phaeopigment, mg m$^{-2}$), surface salinity, and bottom depth. Integrated pigment represents food availability, while surface salinity is an indicator of sea ice formation and melting, and the presence of glacial meltwater nearshore, which affects the type of phytoplankton present in the water column. Relationships between zooplankton and bottom depth potentially indicate concentration of organisms nearshore or in association with Marguerite Trough, or deep shelf depressions. Fall integrated pigment concentrations in Marguerite Bay ranged between 9.6 - 33 mg m$^{-2}$ in 2001, and 72.4 - 236 mg m$^{-2}$ in 2002. Maximum pigment concentrations were located in Laubeuf Fjord during both years, while lowest values were observed in the in George VI Sound during fall 2001, in Crystal Sound during fall 2002, as well as in the vicinity of Alexander Island during both years. Even though pigment concentrations were lowest in Crystal Sound during 2002, values were more than double those observed in the same area during the previous year. Surface salinity values were similar between years and ranged between 33.04 - and 33.55. The shallowest stations had bottom depths of ~ 300 - 400 m and were located near the coast of Alexander Island, the northern edge of Marguerite Trough in central Marguerite Bay, southern Adelaide Island, and inner Marguerite Bay, while the deepest areas, with bottom depths > 800 m, were in northern George VI Sound, Lazarev Bay, and a location in the vicinity of Alexander Island.

Overall, there were no clear associations between the horizontal distribution of zooplankton (ind m$^{-2}$) and concurrent environmental variables, including vertically
integrated pigment concentrations (chlorophyll + phaeopigment, mg m⁻²), surface salinity, and bottom depth (Table 4.5). In general, only a few copepods showed a significant correlation with environmental variables. During 2001, *Ctenocalanus* spp. showed a positive correlation with pigment concentrations, *Oithona* spp. showed a negative relationship, and *G. tenuispinus* was positively correlated with bottom depth. During fall 2002, *Paraeuchaeta* spp. showed a positive correlation with pigment concentrations, while *C. acutus* and *Oncaea* spp. were positively correlated with bottom depth. Only two other zooplankton taxa showed significant relationships with pigment concentrations: *T. macrura* in 2001 and mysids in 2002. Only two groups, *E. superba* and the copepod *R. gigas*, showed significant relationships with surface salinity, and these were only observed during 2002. Of the 138 relationships examined between zooplankton abundance and environmental variables, 11 were significant, which is only slightly higher than the number that would be expected with 95% confidence if all relationships were statistically insignificant (i.e., seven).

At many locations there was a negative trend between the vertical distribution of macrozooplankton (i.e., euphausiids, amphipods, and mysids) and gradients in temperature, salinity, and density (examples in Appendix 3). *E. superba* was located either above or below the thermocline/pycnocline in 92% of the net hauls in which the species was present, while corresponding values for *E. crystallorophias* and *T. macrura* were 77% and 60% respectively (Table 4.6). However, euphasuiids coincided with physical gradients at 8 - 23% of the locations sampled (examples in Appendix 4). Most amphipods and mysids were located primarily deeper than the thermocline/pycnocline, with only smaller fractions present at the depth of the temperature/density gradient. In
general, no patterns were observed in the vertical distribution of zooplankton in relation to fall pigment concentrations.

Table 4.5. Spearman rank order correlations between integrated abundance of zooplankton (ind m$^{-2}$) and vertically integrated pigment concentrations (chlorophyll + phaeopigment; mg m$^{-2}$), salinity at 10 m ($S_{10}$), and bottom depth (bottom $Z$). Significant correlations ($p < 0.05$) are in bold.

<table>
<thead>
<tr>
<th>Taxonomic group</th>
<th>Fall 2001</th>
<th></th>
<th></th>
<th>Fall 2002</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Pigment</td>
<td>$S_{10}$</td>
<td>$Z$</td>
<td>Pigment</td>
<td>$S_{10}$</td>
<td>$Z$</td>
</tr>
<tr>
<td><strong>Copepods</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. acutus</em></td>
<td>0.415</td>
<td>-0.021</td>
<td>-0.147</td>
<td>0.262</td>
<td>-0.161</td>
<td><strong>0.630</strong></td>
</tr>
<tr>
<td><em>C. propinquus</em></td>
<td>0.085</td>
<td>0.385</td>
<td>-0.403</td>
<td>0.582</td>
<td>0.301</td>
<td>-0.144</td>
</tr>
<tr>
<td><em>Ctenocalanus</em> spp.</td>
<td><strong>0.799</strong></td>
<td>-0.063</td>
<td>0.028</td>
<td>0.000</td>
<td>0.210</td>
<td>0.256</td>
</tr>
<tr>
<td><em>G. tenuispinus</em></td>
<td>-0.257</td>
<td>0.211</td>
<td><strong>0.641</strong></td>
<td>0.077</td>
<td>-0.500</td>
<td>0.004</td>
</tr>
<tr>
<td><em>M. gerlachei</em></td>
<td>0.427</td>
<td>-0.126</td>
<td>-0.077</td>
<td>0.211</td>
<td>0.049</td>
<td>0.441</td>
</tr>
<tr>
<td><em>Oithona</em> spp.</td>
<td><strong>-0.701</strong></td>
<td>0.266</td>
<td>0.182</td>
<td>0.143</td>
<td>0.189</td>
<td>0.014</td>
</tr>
<tr>
<td><em>Oncaea</em> spp.</td>
<td>-0.463</td>
<td>-0.161</td>
<td>0.280</td>
<td>0.152</td>
<td>-0.378</td>
<td><strong>0.635</strong></td>
</tr>
<tr>
<td><em>Paraeuchaeta</em> spp.</td>
<td>-0.018</td>
<td>0.154</td>
<td>-0.336</td>
<td><strong>0.743</strong></td>
<td>-0.223</td>
<td>0.217</td>
</tr>
<tr>
<td><em>R. gigas</em></td>
<td>-0.300</td>
<td>0.413</td>
<td>-0.039</td>
<td>0.536</td>
<td><strong>0.609</strong></td>
<td>0.004</td>
</tr>
<tr>
<td><em>S. minor</em></td>
<td>-0.173</td>
<td>0.295</td>
<td>-0.033</td>
<td>-0.081</td>
<td>0.037</td>
<td>0.221</td>
</tr>
<tr>
<td>TOTAL copepods</td>
<td>0.543</td>
<td>-0.049</td>
<td>-0.147</td>
<td>0.397</td>
<td>0.203</td>
<td>0.497</td>
</tr>
<tr>
<td><strong>Ostracods</strong></td>
<td>0.341</td>
<td>-0.126</td>
<td>-0.049</td>
<td>0.379</td>
<td>-0.132</td>
<td>0.382</td>
</tr>
<tr>
<td><strong>Euphausiids</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>E. crystallorophias</em></td>
<td>-0.355</td>
<td>-0.487</td>
<td>0.249</td>
<td>0.110</td>
<td><strong>-0.657</strong></td>
<td>-0.060</td>
</tr>
<tr>
<td><em>E. superba</em></td>
<td>0.220</td>
<td>-0.273</td>
<td>-0.427</td>
<td>0.110</td>
<td>-0.582</td>
<td>-0.109</td>
</tr>
<tr>
<td><em>T. macrura</em></td>
<td><strong>0.707</strong></td>
<td>-0.133</td>
<td>-0.287</td>
<td>-0.633</td>
<td>0.217</td>
<td>0.193</td>
</tr>
<tr>
<td><strong>Amphipods</strong></td>
<td>0.628</td>
<td>-0.035</td>
<td>-0.517</td>
<td>0.008</td>
<td>-0.356</td>
<td>-0.199</td>
</tr>
<tr>
<td><strong>Mysids</strong></td>
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<td>-0.028</td>
<td>-0.028</td>
<td><strong>0.802</strong></td>
<td>-0.347</td>
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</tr>
<tr>
<td><strong>Medusae</strong></td>
<td>-0.300</td>
<td>0.099</td>
<td>0.218</td>
<td>0.206</td>
<td>-0.250</td>
<td>0.411</td>
</tr>
<tr>
<td><strong>Polychaetes</strong></td>
<td>0.122</td>
<td>-0.553</td>
<td>0.039</td>
<td>0.649</td>
<td>-0.189</td>
<td>0.466</td>
</tr>
<tr>
<td><strong>Pteropods</strong></td>
<td>-0.311</td>
<td>0.434</td>
<td>-0.301</td>
<td>0.253</td>
<td><strong>0.580</strong></td>
<td>0.311</td>
</tr>
<tr>
<td><strong>Chaetognaths</strong></td>
<td>-0.280</td>
<td>0.203</td>
<td>-0.028</td>
<td>-0.084</td>
<td>0.308</td>
<td>-0.045</td>
</tr>
<tr>
<td><strong>Appendicularians</strong></td>
<td>-0.165</td>
<td>0.340</td>
<td>-0.119</td>
<td>0.556</td>
<td>0.104</td>
<td>-0.025</td>
</tr>
<tr>
<td><strong>Salps</strong></td>
<td>0.009</td>
<td>0.431</td>
<td>-0.211</td>
<td>-0.055</td>
<td>-0.257</td>
<td>0.354</td>
</tr>
</tbody>
</table>
Table 4.6. Percentage of net hauls in which macrozooplankton were located primarily shallower, deeper, or at the same depth as the thermocline/pycnocline. Percentage of net hauls with widespread vertical distributions are also indicated. *n* is the total number of net hauls in which a taxon was present during 2001 and 2002 combined. A total of seven net hauls were excluded from the calculations due to a vertically uniform water column (four net hauls in 2001) or lack of concurrent CTD data (three net hauls in 2002).

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Shallower (%)</th>
<th>Deeper (%)</th>
<th>Same depth (%)</th>
<th>Widespread (%)</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Euphausiids</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>E. crystallorophias</em></td>
<td>54</td>
<td>23</td>
<td>23</td>
<td>0</td>
<td>13</td>
</tr>
<tr>
<td><em>E. superba</em></td>
<td>58</td>
<td>33</td>
<td>8</td>
<td>0</td>
<td>12</td>
</tr>
<tr>
<td><em>T. macrura</em></td>
<td>33</td>
<td>27</td>
<td>13</td>
<td>27</td>
<td>15</td>
</tr>
<tr>
<td>Amphipods</td>
<td>19</td>
<td>56</td>
<td>6</td>
<td>19</td>
<td>16</td>
</tr>
<tr>
<td>Mysids</td>
<td>0</td>
<td>89</td>
<td>11</td>
<td>0</td>
<td>9</td>
</tr>
</tbody>
</table>

**Summer Chlorophyll Concentrations and Krill Recruitment**

Surface chlorophyll concentrations in the Bellingshausen Sea and Marguerite Bay during the spring and summer influenced the abundance and composition of the zooplankton during fall (Fig. 4.11a). In oceanic waters of the Bellingshausen Sea (Fig. 4.11b and c), chlorophyll concentrations were above average during austral spring-summer 2000/2001 (November - January). As the summer progressed, phytoplankton blooms moved onshore and above average chlorophyll concentrations were observed in the coastal Bellingshausen Sea during January - March 2001 (Fig. 4.11d). During the 2001/2002 season, conditions offshore were average between November and January (Fig. 4.11b and c), while in coastal waters, above normal chlorophyll concentrations were primarily observed by late February (Fig. 4.11d). Within Marguerite Bay, the interannual differences were even more striking, with extremely high chlorophyll concentrations during austral summer - fall 2001 and below average conditions in 2002 (Fig. 4.11e - f).
There was a strong correlation (Spearman R = 0.81, p < 0.05) between geometric mean chlorophyll concentrations in the Bellingshausen Sea during November and summer recruitment indices for *E. superba* previously reported for the western Antarctic Peninsula region (Fig. 4.12a). Geometric mean chlorophyll concentrations for November 1997 - 2004 in oceanic waters of the Bellingshausen Sea were highest during the spring - summer 2000/2001 season (n = 8), coincident with the highest recruitment index (R₁) observed for *E. superba* during all years for which data are available (1997/1998 - 2002/2003) (Fig. 4.12b). Relatively low chlorophyll concentrations were registered during November 1997, 1998, and 2002, when low recruitment values also were recorded.
Figure 4.11. (a) Location of the subregions analyzed for median SeaWiFS chlorophyll concentrations (chl, mg m$^{-3}$) in (b, c) oceanic and (d) coastal waters of the Bellingshausen Sea, (e) northern, and (f) southern Marguerite Bay in spring/summer 2000/2001 (grey) and 2001/2002 (black). A climatology for spring/summer 1997/1998 - 2003/2004 is represented by the red line and corresponds to the median chlorophyll in each subregion for the seven seasons analyzed.
Figure 4.11. (Continued)
Figure 4.12. (a) Spearman R correlation between geometric mean chlorophyll concentrations (Chl, mg m$^{-3}$) in oceanic waters of the Bellingshausen Sea during November 1997 - 2004 and recruitment of *E. superba* (R$_1$) in waters west of the Antarctic Peninsula, for the period 1997/1998 - 2002/2003; n = 6; R = 0.81; p < 0.05). (b) Time series of geometric mean chlorophyll concentration in oceanic waters of the Bellingshausen Sea during November 1997 - 2004 (grey bars) and recruitment of *E. superba* (R$_1$) (black circles) in waters west of Antarctic Peninsula. Chlorophyll estimated as geometric mean SeaWiFS chlorophyll concentration during late November in subregion 2 of Fig. 4.11a. Recruitment indices (R$_1$) for 1997/1998 - 1999/2000 from Siegel et al. (2002); for 2000/2001 from Siegel et al. (2003); and for 2001/2002 and 2002/2003 from the Palmer LTER DataZoo database (data provided by L. Quetin and R. Ross).
DISCUSSION

Composition and Abundance of Zooplankton in Marguerite Bay

Total zooplankton abundances in the WAP, and Marguerite Bay in particular, are generally higher than those reported for other areas of the Southern Ocean. Deibel and Daly (2007) summarized zooplankton abundance data for continental shelf regions around Antarctica and report that values in Marguerite Bay and Crocker Passage are one or two orders of magnitude higher than in any other area considered, including the Weddell and Ross seas. Copepods numerically dominated the zooplankton community of Marguerite Bay during fall, with a mean abundance of 24,362 ind m$^{-2}$ in 2001, and values up to 44,135 ind m$^{-2}$ in Laubeuf Fjord. Also during SO GLOBEC, Ashjian et al. (2008) investigated zooplankton on the outer continental shelf of Marguerite Bay using Video Plankton Recorder (VPR) data, and estimated copepod mean integrated abundances of 2,832 ± 2,983 ind m$^{-2}$ for fall 2001, only ~ 10% of the estimates reported here for coastal waters in Marguerite Bay. In comparison, near South Georgia in the Scotia Sea, total copepod abundances during summer 1994 - 1996 were up to an order of magnitude higher than those calculated here for fall, with median values ranging between 66,684 and 235,793 ind m$^{-2}$ for the upper 200 m of the water column (Atkinson et al., 1999).

Copepod species composition in Marguerite Bay was comparable to that described for other Antarctic areas, including waters west of the Antarctic Peninsula (e.g., Hopkins, 1985; Schnack-Schiel and Mujica, 1994), the Bellingshausen Sea (Atkinson, 1995; Siegel and Harm, 1996), and western Weddell Sea (Hopkins and Torres, 1988). However, during SO GLOBEC, $C. acutus$, $M. gerlachei$, $Ctenocalanus$ spp., and $C.$
propinquus dominated total copepod abundance and, the smaller Oncaea spp. and Oithona spp. did not represent a major fraction of the community, in contrast to previous results. A study in Crocker Passage, on the northern Antarctic Peninsula shelf, reported a similar fall copepod species composition, but observed that Oncaea curvata comprised over half of the total abundance (Hopkins, 1985). Other results from the WAP also indicate that these smaller copepods generally outnumber larger species (Schnack-Schiel and Mujica, 1994; Cabal et al., 2002). In contrast to the relatively finer mesh used during these previous studies (162 - 200 μm), zooplankton were sampled with 333 μm mesh during SO GLOBEC; thus, it is possible that abundances of smaller cyclopods, including Oncaea spp. (0.6 - 1.1 mm in length) and Oithona spp. (0.7 - 1.2 mm), were underestimated.

T. macrura was the most abundant euphausiid in 2001, with integrated abundances up to 266 ind m$^{-2}$. These values are higher than maximum integrated abundances of 78 ind m$^{-2}$ reported during spring 1989 for Gerlache Strait, at the northern end of the WAP (Nordhausen, 1994). Mean abundances of T. macrura during fall in Marguerite Bay (2001: 45.9 ind m$^{-2}$; 2002: 16.2 ind m$^{-2}$) also were higher than average spring values in the Weddell Sea, which ranged between 0.75 and 2.98 ind m$^{-2}$ in the upper 200 m (Donnelly et al., 2006). E. superba is often a major fraction of the macrozooplankton community, comprising up to 95% of the larger zooplankton in waters along the northern WAP (Lancraft et al., 2004). This species was observed at almost every location surveyed during the present study; however, abundances did not exceed those of other euphausiids and macrozooplankton groups, particularly during 2001. The distribution of E. superba is patchy, with post larval individuals usually aggregated in
compact and dense swarms that can often be undersampled by plankton nets (Wiebe et al., 2004); thus, the relatively low abundances reported here should be interpreted with caution. Densities of *E. superba* estimated from acoustic surveys during the fall survey cruises in Laubeuf Fjord and Crystal Sound were approximately an order of magnitude higher than those from net samples in the same area, and the difference was attributed, in part, to krill net avoidance (Lawson et al., 2008). *T. macrura* forms more diffuse aggregations and is more evenly distributed throughout the water column, which allows for more accurate abundance estimates from net data. Despite being likely undersampled, *E. superba* still comprised a major fraction of the fall zooplankton biomass in Marguerite Bay (see estimates below). Mean integrated abundances of juvenile and adult *E. superba* in Marguerite Bay were 6.22 and 14.8 ind m$^{-2}$ in 2001 and 2002, respectively, with maximum values up to 117 ind m$^{-2}$ in Crystal Sound during 2002. These abundances are in the same order of magnitude as values reported for other areas of the WAP. In a review of data for Elephant Island between 1977 and 2004, Siegel (2005) reports abundances ranging between 1.4 and 336 ind m$^{-2}$ (median for all years = 9.1 ind m$^{-2}$), while Lancraft et al. (2004) estimated fall abundances of 788 ind m$^{-2}$ for the upper 200 m of the water column in Crocker Passage.

Although *T. macrura* may at times outnumber *E. superba*, the latter is generally more important in terms of biomass. For the present study, biomass data are not available; however, some simple calculations can provide useful estimates of interspecific biomass differences. During 2001, the overall geometric mean abundance of *T. macrura* in Marguerite Bay was 45.9 ind m$^{-2}$, with most individuals at 9 - 10 mm TL (Fig. 4.5). In the case of *E. superba*, the average abundance was 6.22 ind m$^{-2}$ during
the same year, with a mode at 51 mm TL (Fig. 4.4). The mean biomass for *T. macrura* in 2001 was 0.68 g WW m\(^{-2}\), while the corresponding value for *E. superba* was 9.11 g WW m\(^{-2}\), based on the length frequency distributions for both species during fall 2001 and a length-wet weight relationship from Ashjian et al. (2004) (wet weight = 0.0054 x Length\(^{3.214}\)). These biomass estimates are within the range of values reported for euphausiids in other Antarctic areas. In the western Antarctic Peninsula shelf region, mean fall biomass of krill estimated from acoustic data was 12 g m\(^{-2}\) (Lascara et al., 1999), whereas summer estimates for krill around Elephant Island in 1978 - 2004 ranged between 0.76 and 75.2 g m\(^{-2}\) (Siegel, 2005). In addition, Voronina (1998) summarized published biomass estimates for *E. superba* from plankton nets in different areas of the northern WAP and reported values generally ranging between 0.2 and 54 g WW m\(^{-2}\), although biomass at Crocker Passage during fall 1983 reached 229 g WW m\(^{-2}\). In the Weddell Sea, average spring biomass for *E. superba* ranged from 0.54 g WW m\(^{-2}\) in open waters, to 1.2 g WW m\(^{-2}\) in the vicinity of the ice edge, while corresponding values for *T. macrura* were 0.57 and 1.4 g WW m\(^{-2}\), respectively (Donnelly et al., 2006). Despite the rapid population response to elevated food concentrations and numerical dominance of *T. macrura* during fall 2001, *E. superba* was still the dominant euphausiid in terms of biomass.

Other numerically important taxa observed in Marguerite Bay during fall 2001 and 2002, such as ostracods, pteropods, polychaetes, and chaetognaths, are common members of the zooplankton of the WAP (Schnack-Schiel and Mujica, 1994; Siegel and Harm, 1996). Pteropods and ostracods constituted the most abundant non-copepod zooplankton in the study area during both years analyzed. Pteropod abundances recorded
during this study did not vary interannually, and are similar to other SO GLOBEC values reported for the outer shelf off Marguerite Bay, which reached ~ 1200 ind m\(^{-2}\) in the vicinity of Marguerite Trough (Ashjian et al., 2008). Small pteropods (< 15 mm), which dominated numerically in Marguerite Bay with values up to 2,119 ind m\(^{-2}\), are mostly herbivores (Hopkins, 1985). Ostracods abundances reached 4,728 ind m\(^{-2}\) in southern Laubeuf Fjord during 2001, which comprised ~ 76% of the non-copepod zooplankton at this location. Ostracods are generally omnivores and feed primarily on debris of \textit{E. superba}, phytoplankton, and copepods (Hopkins, 1985). To the author’s knowledge, other ostracod integrated abundances have not been reported for the Marguerite Bay area, and VPR results indicated that they were not a dominant component of the zooplankton of the mid and outer shelf of Marguerite Bay (C. Ashjian, \textit{pers. comm.}). A study in the Weddell Sea observed that ostracods comprised 5.4% of total zooplankton biomass under the ice, and 2.6% in open waters (Hopkins and Torres, 1988).

Chaetognaths were more abundant in Marguerite Bay than in other Antarctic areas, with mean integrated abundances of 57 and 31.3 ind m\(^{-2}\) in 2001 and 2002 respectively, although values reached 687 ind m\(^{-2}\) in the vicinity of Alexander Island during 2001. Donnelly et al. (2006) report mean values of 1.75 ind m\(^{-2}\) for the Weddell Sea, while Lancraft et al. (2004) estimated mean integrated abundances of 6.37 ind m\(^{-2}\) for Crocker Passage during fall. During SO GLOBEC, chaetognaths comprised, on average, 8% of the non-copepod zooplankton during 2001, with values up to 42% in Lazarev Bay, and 23% around Alexander Island. The mean percent composition reported here agrees well with values from the Weddell Sea ranging between 3.8 and 29.3% (Hopkins and Torres, 1988; Donnelly et al., 2006). Chaetognaths are predators and feed
mostly on copepods, although the larger species also have been observed to prey on mysids and amphipods (Hopkins, 1985). Thus, high abundances of chaetognaths in copepod-rich waters near Alexander Island are not surprising.

Polychaetes were more abundant in fall 2001 relative to 2002. This group includes mostly herbivorous species in Antarctic waters (Hopkins, 1985) and, thus, higher densities in fall 2001, when chlorophyll concentrations were higher, are expected. Vertically integrated abundances reported for the outer shelf off Marguerite Bay reached \( \sim 2,300 \text{ ind m}^{-2} \) west of Alexander Island in 2001 (Ashjian et al., 2008), more than double the maximum values of 1,110 ind m\(^{-2}\) estimated here in coastal Marguerite Bay during the same year.

**Variability in Euphausiid Life History Strategies**

Variability in life history strategies between euphasuiid species resulted in interannual differences in species percent composition. During the chlorophyll-rich 2001 season, the ubiquitous *T. macrura* dominated the euphausiid community, followed by *E. superba*, and *E. crystallorophias*. The numerical dominance of *T. macrura* has been previously reported as well for other areas of the Southern Ocean (e.g., Atkinson and Peck, 1988; Mujica, 1989). There exist marked differences in the timing of the onset of reproduction between species of Antarctic euphasiids. *T. macrura* is the first species to start reproducing, with spawning as early as September (Makarov, 1979). Later in spring, *E. crystallorophias* reproduction begins, followed by *E. superba*, which spawns mainly between November and March (Marr, 1962).

Most individuals of *T. macrura* observed during fall 2001 were juveniles between
8 - 11 mm TL (87% total) (Fig. 4.5). *T. macrura* has a more rapid development than the *Euphausia* species. Although information on larval growth rates is available for *T. macrura* (Nordhasuen, 1992, Siegel, 1987), juvenile development has not been described. Nordhausen (1992) reports that it takes 90 days for calyptopis II (C2) to develop into the last larval stage, furcilia VI (F6). In addition, Makarov (1979) estimates that it takes 15 - 20 days for eggs to change into C2, indicating a total larval development time of 105 - 110 days. Siegel (1987) estimated age and growth of *T. macrura* in the Weddell Sea from length-frequency data and suggested that larvae can first develop into juveniles at 8 mm in length, during the second half of their first year of life. If reproduction of *T. macrura* along the western Antarctic Peninsula starts in September - October, individuals observed during fall could be up to seven or eight months old. Development estimates suggest that the *T. macrura* juveniles observed during fall 2001 originated from a reproductive event(s) during spring-summer 2000/2001 (age-class 0+), when chlorophyll concentrations were above climatology values.

Development of *E. superba* and *E. crystallorophias* is slower and, therefore, populations take longer to respond to environmental changes. In contrast to the 105 - 110 days estimated for larval development of *T. macrura*, laboratory experiments indicate that *E. superba* can develop from an egg to F6 in approximately 127 days (Ikeda, 1984), while the larval development time for *E. crystallorophias* is even longer (Ikeda, 1986; Brinton and Townsend, 1991). This longer development coupled with a later onset of reproduction in November - March suggest that juveniles of *E. superba* and *E. crystallorophias* will not be present in the water column of the western Antarctic Peninsula region until the spring of their second year (age class 1+), which is supported
by field observations (Daly 2004; Daly and Zimmerman, 2004). During SO GLOBEC, elevated chlorophyll concentrations in 2001 supported a successful reproduction of *E. superba* and *E. crystallorophias* during spring-summer 2000/2001, as evidenced by the presence of numerous larvae in fall 2001 and juveniles during 2002. Hence, *T. macrura* had a rapid population response to elevated chlorophyll concentrations, demonstrated by the large numbers of juveniles present in fall 2001 (Fig. 4.5), whereas *E. superba* and *E. crystallorophias* showed a slower population response, supported by the scarcity of juveniles of either species during fall 2001 and the high proportion of juveniles present during fall 2002 (Figs 4.4 and 4.6).

*E. superba* females are believed to spawn offshore in the vicinity of the shelf-break (Marr, 1962; Siegel, 1988; 1992). Because eggs are denser than seawater, they sink and hatch at depths of 800 - 1000 m. The young larvae then swim to the surface before turning into the first feeding stage, calyptopis I (Marr, 1962), at which point they need to find food within approximately 10 days or otherwise will not survive (Ross and Quetin, 1986). This life strategy is hypothesized to have several ecological advantages, including preventing the eggs from reaching the seafloor and becoming unviable or eaten by benthic organisms. In addition, the larvae produced at these depths will develop in warmer (> 1 °C) Circumpolar Deep Water (CDW) found below 500 m, thus reducing the risk of predation by epi- and mesopelagic fauna. The developmental ascent described by Marr (1962) for *E. superba* also has been reported for larvae of *T. macrura* (Makarov, 1979). These two species share several ecological characteristics, such as a common widespread distribution, a similar reproductive strategy involving deep hatching of eggs and larval ascent, and a primarily herbivorous larval phase. Their similar life strategies
would indicate that these euphausiids are strong competitors; however, it has been suggested that the two month separation between the onset of reproduction is an adaptation to avoid competition for food between their larvae (Makarov, 1979). In addition, during the postlarval stages, when both species coexist, *E. superba* are mainly herbivores during spring and summer, while adult *T. macrura* have been described as omnivores, feeding mostly on large copepods, such as *C. acutus* and *M. gerlachei* (Hopkins, 1985). Additionally, their depths of maximum abundances in the water column generally do not overlap. In summary, although *E. superba* and *T. macrura* share a common distribution and similar life history strategies, they have developed individual adaptations to minimize competition between them, allowing them to coexist as widespread, successful, Antarctic species.

**Summer Chlorophyll and Zooplankton Population Response**

Interannual variability in chlorophyll concentrations strongly influences zooplankton populations. Population responses for *T. macrura*, *E. crystallorophias*, and *E. superba* were described above. In addition, the influence of spring/summer chlorophyll concentrations is demonstrated by the strong correlation observed between November chlorophyll concentrations in the Bellingshausen Sea and recruitment of *E. superba* along the WAP during the following year. Previous studies reported correlations between sea ice extent during winter and successful juvenile recruitment of *E. superba* during the following spring (Kawaguchi and Satake, 1994; Siegel and Loeb, 1995; Hewitt et al., 2003). Sea ice biota on the undersurface of sea ice constitutes an alternative food source for overwintering larval krill, while sea ice provides refuge from predators, which
further reduces winter larval mortality (Daly 1990; Daly and Macaulay, 1991). During SO GLOBEC, there were interannual differences in the extent and timing of the advance and retreat of sea ice (Chapter 3; Parkinson, 2002; Marrari et al., 2008); however, winter sea ice conditions were similar in 2001 and 2002, and sea ice biota concentrations were low at the ice-water interface during both years (0.05 - 0.07 µg l⁻¹) (Daly, 2004). Despite similar winter sea ice, krill recruitment showed marked interannual differences, suggesting that other processes influence recruitment of *E. superba* in this area. It is here hypothesized that the early and persistent availability of phytoplankton in offshore waters of the Bellingshausen Sea supports early and repeated reproductive events during spring and summer and results in a high reproductive output. In addition, high concentrations of phytoplankton in offshore and coastal areas during summer and fall will lead to faster growth and development of larvae, which will be in better condition to survive overwinter. Low food conditions or the late onset of blooms will lead to late and/or poor reproduction and smaller/weaker larvae which may not be able survive overwinter. Although phytoplankton blooms in the Bellingshausen Sea and Marguerite Bay were spatially variable, spring chlorophyll concentrations exceeded minimum values (1 - 5 mg m⁻³) required to initiate reproduction of *E. superba* (Ross and Quetin, 1986) during both SO GLOBEC years. It is the variability in the timing and persistence of these blooms, rather than the absolute chlorophyll concentrations, what will influence the reproduction and recruitment success of euphausiids.

The composition of the copepod community also showed marked interannual differences, which appear to be related to variability in food availability. During the chlorophyll-rich 2001 season, *C. acutus* dominated, comprising 52% of the total
copepods. *C. acutus* has been described as the only true herbivore in Antarctic waters (Conover and Huntley, 1991; Atkinson, 1998), feeding mainly on phytoplankton during spring and summer. This species spawns over a short time and produces one distinct cohort. Maximum densities can be observed in the surface layer during summer, when the population reaches stage copepodid V (C5). In areas of deep bathymetry, *C. acutus* undergoes a seasonal ontogenetic migration, moving to deep waters during the fall and winter, where individuals do not feed (Schnack-Schiel and Mujica, 1994), whereas in shallower coastal areas, *C. acutus* moves to intermediate depths. *M. gerlachei*, the second numerically dominant species during 2001 and most abundant copepod in 2002, is primarily omnivorous and its success does not rely heavily on high phytoplankton concentrations. This species starts to spawn in December and continues during summer. *M. gerlachei* does not undergo a seasonal ontogenetic migration and is more widely distributed throughout the water column (Atkinson and Peck, 1988; Schnack-Schiel and Hagen, 1995). The elevated phytoplankton concentrations observed in Marguerite Bay throughout the summer of 2001 likely favored the reproduction of *C. acutus*, which reached relatively high abundances and dominated the copepod community during fall. Lower chlorophyll concentrations during 2002 resulted in lower abundances of copepods. The copepod community was dominated by omnivorous species, such as *M. gerlachei* and *Ctenocalanus* spp., which together accounted for ~60% of total copepods, whereas *C. acutus* had an 85% population reduction relative to 2001.

*Controls on Zooplankton Spatial Patterns in Marguerite Bay*

Copepods and macrozooplankton showed opposite horizontal distributions in
Marguerite Bay, with higher abundances of macrozooplankton in the northern sectors of the study area, and greatest densities of copepods in southern areas. These distinct distributions are likely the result of a combination of circulation features and seasonal predation. The general circulation in Marguerite Bay (Fig. 1.2) involves a coastal current (APCC) that enters Marguerite Bay at the northern end, flows along the coast in a clockwise direction (Moffat et al., 2008), and exits Marguerite Bay along the outer coast of Alexander Island. In addition, drifter data revealed a clockwise gyre-like circulation in the central areas of Marguerite Bay during fall 2001 and 2002 (Beardsley et al., 2004), while ADCP results showed the presence of two eddies in George VI Sound during fall 2001 (Dorland and Zhou, 2008). These features, in combination with the general circulation of the APCC, contribute to creating a favorable retention habitat for zooplankton in Marguerite Bay. These circulation features can transport zooplankton within coastal Marguerite Bay, particularly smaller taxa such as copepods, which are unable to swim against currents. The northern sectors of the study area, such as southern Crystal Sound, had a more sluggish flow than that observed for the APCC (Zhou et al., 2004). Given the capabilities of euphasiids to swim at cruising speeds of 10 - 15 cm sec\(^{-1}\) (Hamner et al., 1983; Zhou and Dorland, 2004), it is likely that these larger organisms are able to maintain their distributions preferentially in these quieter northern areas, resulting in the higher abundances observed relative to the southern sectors. In a study from the Crystal Sound area using Acoustic Doppler Current Profiler (ADCP) data, Zhou and Dorland (2004) demonstrate that the swimming capabilities of *E. superba* can determine the maintenance of aggregations in the mesoscale circulation field of the area.

The effects of predation are cumulative over the summer productive period and
may have contributed to the different spatial distributions observed between copepods and euphausiids in Marguerite Bay during fall. Predation during fall was likely not strong enough to significantly reduce copepod abundances in the northern sectors, particularly given that the much of the euphausiid and copepod populations occurred at different depths in the water column. A negative correlation between abundances of copepods and euphausiids was reported for summer in waters around South Georgia, where predation pressure of euphausiids was found to determine, at least in part, copepod distributions (Atkinson et al., 1999). In summary, the combined effects of currents and retention features, seasonal predation, and behavior of macrozooplankton, likely determined the distinct spatial distributions observed for these taxa during fall in Marguerite Bay.

Vertical distribution of euphausiids and other zooplankton, including copepods, amphipods and mysids, rarely overlapped throughout Marguerite Bay during 2001. All euphausiids were consistently shallower than the most abundant copepods species, *C. acutus* and *M. gerlachei*, and these vertical distributions are consistent with other results for euphausiids of the continental shelf of Marguerite Bay (e.g., Lawson et al., 2008; Ashjian et al., 2008). Considering that the fall 2001 cruise took place during May - June, after *C. acutus* has normally started its seasonal descent into deeper waters, the vertical separation of the two groups could be attributed to differing winter behaviors. However, the omnivore *M. gerlachei*, which also comprised a major fraction of total copepods, does not undergo seasonal vertical migrations and is known to feed throughout the year. Thus, the deep distribution of this species relative to euphausiids could be the result of accumulated predation pressure. In contrast to the vertical depth partitioning observed in
2001, euphasiids and copepods had an overlapping distribution during 2002. The overall shallower distribution of copepods in 2002 could have resulted from a combination of factors, including (1) an earlier fall cruise (April - May) relative to 2001 (May - June), (2) a smaller contribution of *C. acutus* to the total copepod abundance with higher proportions of surface-dwelling species such as *Ctenocalanus* spp and *Oithona* spp., and (3) reduced predation pressure during fall 2002, a consequence of *E. superba* not feeding on copepods (K. Daly, *pers. comm.*), as well as lower densities of other omnivorous and carnivorous zooplankton, such as *T. macrura*, chaetognaths and medusae.

Studies of mysids from the Southern Ocean are few; however, Brandt et al. (1998) described 37 species of Antarctic mysids, 19 of which are endemic, and reviewed information on their biogeography and vertical distributions. Mysids are generally hyperbenthic or bathypelagic, and prefer an omnivorous diet, feeding on phytoplankton and a variety of zooplankton, including copepods, coelenterates, and euphausiid molts (Hopkins, 1985). During our study, mysids were located consistently deeper than euphausiids and amphipods, with some individuals recorded at depths of up to 800 m, the maximum depth sampled during SO GLOBEC. These distributions are consistent with previous coastal studies, while in deep-waters, Antarctic species have been recorded at depths of up to 4500 m (Torres and Hopkins, 1988; Brandt et al, 1998). Although depths in Marguerite Bay reach ~ 1,600 m at a few locations, the mean depth of the area is ~ 400 m (Bolmer et al., 2004) (Fig. 1.1). In deeper areas of the WAP, such as the vicinity of the shelf break, vertical distributions of mysids may extend to greater depths than those reported herein.
Amphipods were located at intermediate depths between euphausiids and mysids in 2001, but were shallower than both groups during 2002. Although data on the species composition of the amphipod community during SO GLOBEC are not currently available, the distinct vertical distributions observed could be related to interannual variability in the percent contribution of different species. For example, in 2001 deeper living large gammarid amphipods appear to have dominated, while smaller hyperiid amphipods, such as *Themisto gaudichaudii*, which are known to inhabit shallower waters (Hopkins, 1985; Torres and Hopkins, 1988), may have been more important in 2002.

**Relationship between zooplankton and fall environmental parameters**

There were no clear trends between zooplankton spatial distributions and fall environmental variables in Marguerite Bay. The positive correlation observed in 2001 between pigment concentrations and abundances of *Ctenocalanus* spp. could be the result of the mainly herbivorous diet of this species (Hopkins, 1985); however, no other herbivores had positive correlations with this parameter. The positive relationship between the carnivore *Paraeuchaeta* spp. and pigment concentrations during 2002 is even less evident. *Paraeuchaeta* spp. are raptorial predators, feeding primarily on smaller copepods, such as *Oncaea* spp. and copepodites of *M. gerlachei* (Hopkins 1985), which are omnivores and may ingest significant amounts of phytoplankton. However, these prey species did not show positive relationships with pigment concentrations. Moreover, only a weak positive trend (not statistically significant) was observed between *Paraeuchaeta* spp. and their main prey, *Oncaea* spp. (r = 0.39; p = 0.216) and *M. gerlachei* (r = 0.32; p = 0.308) during 2002. The overall lack of relationship between
pigment concentrations and grazers is likely due to the fact that herbivorous/omnivorous copepods may not have been feeding on phytoplankton during mid to late fall. Among the macrozooplankton, only the omnivores *T. macrura* and mysids showed positive correlations with pigment concentrations in 2001 and 2002 respectively, but these relationships were not consistent interannually.

Relationships between copepod abundances and bottom depth are consistent for some copepod species (Table 4.4). For example, *G. tenuispinus*, *C. acutus*, and *Oncaea* spp. were classified as deep species during both years, and also showed a significant tendency for higher abundances at locations of deeper bathymetry, such as those in the vicinity of Marguerite Trough and southern Laubeuf Fjord (Fig. 1.3).

**Summary**

The variability observed in total abundance and percent composition of the zooplankton of Marguerite Bay during fall can be linked directly to the contrasting environmental conditions that prevailed during the preceding spring-summer seasons. During spring-summer 2000/2001 a combination of warmer sea surface temperatures, higher than normal chlorophyll concentrations, and low sea ice cover resulted in a favorable environment for zooplankton reproduction and larval growth, which led to the distinctive composition and overall higher abundances observed during fall 2001. Above average concentrations of chlorophyll in many areas, including the Bellingshausen Sea and Marguerite Bay, led to high concentrations of copepods, juvenile *T. macrura*, and larval *Euphausia* spp. during fall 2001, and subsequent elevated numbers of juvenile and adult *E. superba* and *E. crystallorophias* in fall 2002. On the other hand, lower surface
temperatures, extensive and persistent sea ice cover and, consequently, lower than normal chlorophyll concentrations during spring-summer 2001/2002, resulted in lower plankton abundances in Marguerite Bay during fall 2002, particularly copepods, larval euphausiids, herbivorous macrozooplankton and juvenile *T. macrura*. For other groups that do not rely heavily on phytoplankton for reproduction and survival, such as ostracods, pteropods, chaetognaths, amphipods and mysids, the interannual environmental variability did not have as large an impact on their populations, which is evidenced by the smaller interannual differences observed in their abundances.
Validated satellite chlorophyll data offer great insight into the temporal and spatial variability of phytoplankton dynamics in remote and difficult to access areas, such as the Southern Ocean. The high resolution and synoptic nature of these datasets are unattainable using traditional ship-based measurements. Validation results confirm that SeaWiFS surface chlorophyll concentrations derived for the Southern Ocean are an accurate measure of \textit{in situ} values between 0.05 - 1.5 mg m\(^{-3}\). These findings contradict previous results that report an underestimation of SeaWiFS chlorophyll in Antarctic waters. The good agreement between SeaWiFS and \textit{in situ} chlorophyll concentrations reported here was based on the use of chlorophyll data determined by HPLC as ground truth, instead of chlorophyll concentrations estimated from fluorescence, which has been shown to introduce significant errors when certain accessory pigments are present in the water column. Because more than 90\% of the Southern Ocean has chlorophyll values in the 0.05 - 1.5 mg m\(^{-3}\) range, it is not necessary to develop an alternative bio-optical algorithm for this region. However, if computer models (e.g., to estimate primary production or eutrophic depth) have been developed using fluorometric methods as input, then the satellite estimates of chlorophyll concentrations will need adjustment to be consistent with these models.

A major finding of this dissertation is the presence of predictable and persistent
phytoplankton blooms in the Bellingshausen Sea and Marguerite Bay areas between 1997 and 2004, and the strong influence that environmental variability in these areas has on the zooplankton community of the WAP. The lack of field sampling in this region may partly explain why the magnitude of phytoplankton aggregations has been previously overlooked. Chlorophyll concentrations in the southern sectors are consistently higher than in any other part of the WAP. The climatology of chlorophyll concentrations for the period between 1997 and 2004 indicated that chlorophyll values were elevated during 2001 in comparison to all other years, particularly in the vicinity of Marguerite Bay.

Zooplankton composition in the vicinity of Marguerite Bay was similar to that reported for previous WAP studies, and included 12 species of copepods, three species of euphausiids, and ten other groups. The observed variability in total abundance and percent composition of the zooplankton during fall was strongly influenced by the contrasting environmental conditions that prevailed during the preceding spring-summer seasons. During spring-summer 2000/2001 warmer sea surface temperatures, higher than normal chlorophyll concentrations, and low sea ice cover created a favorable environment for zooplankton reproduction and larval growth. Above average chlorophyll concentrations in 2001 led to high concentrations of copepods, juvenile *T. macrura*, and larval *Euphausia* spp. during fall 2001, and subsequent elevated abundances of juvenile and adult *E. superba* and *E. crystallorophias* during fall 2002. In contrast, lower surface temperatures, extensive and persistent sea ice cover, and consequently lower than normal chlorophyll concentrations during spring-summer 2001/2002 resulted in lower plankton abundances in Marguerite Bay during fall 2002, particularly copepods, larval euphausiids, herbivorous macrozooplankton and juvenile *T. macrura*. For other groups
that do not rely heavily on phytoplankton for reproduction and survival such as ostracods, pteropods, chaetognaths, amphipods and mysids, environmental variability did not have a large impact on their populations, which is evidenced by the small interannual differences observed in their abundances.

Zooplankton, particularly *E. superba*, originating in the Bellingshausen Sea and southern areas along the WAP may be a significant source for populations found downstream in shelf waters of the northern Antarctic Peninsula, the Scotia Sea, and South Georgia (Fach and Klinck, 2006; Thorpe et al, 2007). Thus, variability in the timing and persistence of phytoplankton in the southern WAP will not only affect local zooplankton and predators, but ultimately may impact the entire ecosystem of the WAP and adjacent Scotia Sea. The pivotal role that environmental variability in Marguerite Bay and the Bellingshausen Sea plays in structuring the zooplankton community is demonstrated by the results presented here. Given the few datasets currently available, and the poor understanding of the processes than control environmental variability in these regions, particularly chlorophyll dynamics, future studies of ecosystem dynamics along the WAP should include the Marguerite Bay and Bellingshausen Sea regions.
REFERENCES CITED


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Appendix 1. Vertical distribution of euphausiids (left column) and copepods (ind m$^{-3}$) (right column) in Marguerite Bay during fall 2001. The black broken line indicates the maximum sampling depth when a net haul was sampled to depths shallower than 500 m.

Crystal Sound

Northern Laubeuf Fjord

Southern Laubeuf Fjord
Appendix 1. (Continued)

Bourgeois Fjord

Neny Fjord

Northern Alexander Island
Appendix 1. (Continued)
Appendix 1. (Continued)

Northern George VI Sound

Southern George VI Sound

Southern George VI Sound
Appendix 2. Vertical distribution of euphausiids (left column) and copepods (ind m$^{-3}$) (right column) in Marguerite Bay during fall 2002. The black broken line indicates the maximum sampling depth when a net haul was sampled to depths shallower than 500 m. Color legend as in Appendix 1.
Appendix 2. (Continued)

Southern Laubeuf Fjord

Northern Alexander Island
Appendix 2. (Continued)
Appendix 2. (Continued)

Marguerite Trough

Southern Adelaide Island

Inner Marguerite Bay
Appendix 3. Vertical distribution of macrozooplankton (ind m$^{-3}$) in Marguerite Bay, in relation to environmental parameters. Pigment (mg m$^{-3}$) represents chlorophyll + phaeopigment concentrations.

Southern Laubeuf Fjord, fall 2001
Appendix 3. (Continued)

Bourgeois Fjord, fall 2001
Appendix 3. (Continued)

Southern George VI Sound, fall 2001
Appendix 3. (Continued)

Crystal Sound, fall 2002
Appendix 3. (Continued)

Southern Laubeuf Fjord, fall 2002
Appendix 3. (Continued)

Inner Marguerite Bay, fall 2002
Appendix 4. Vertical distribution of macrozooplankton (ind m$^{-3}$) in Marguerite Bay, in relation to environmental parameters. Pigment (mg m$^{-3}$) represents chlorophyll + phaeopigment concentrations.

Northern Laubeuf Fjord, fall 2001
Appendix 4. (Continued)

Northern Laubeuf Fjord, fall 2002
ABOUT THE AUTHOR

Marina Marrari received a degree in Biological Sciences from the Universidad Nacional de Mar del Plata, Argentina, in 2001. She did her thesis research at the Zooplankton Laboratory of the Instituto Nacional de Investigación en Desarrollo Pesquero (INIDEP) under the guidance of Dr. María Delia Viñas, and later worked at Scripps Institution of Oceanography for several months in 2002. During this time, Marina participated in numerous research cruises in continental shelf waters off the coast of Argentina and California.

Marina came to the College of Marine Science of the University of South Florida as a Fulbright Scholar in the fall of 2002, and joined the Zooplankton Ecology Laboratory led by Dr. Kendra Daly. While in the Ph.D. program, she participated in several international research meetings and coauthored four publications in peer-reviewed scientific journals.